

Simulated drought with Polyethylene-Glycol (PEG) decreases above-ground performance and increases nodulation in the legume *Medicago lupulina*

Hana Cho^{1§}, Emily Glasgow¹, Valmic Mukund¹, Julia A. Boyle^{1,2}, John R. Stinchcombe¹

¹Department of Ecology and Evolutionary Biology, University of Toronto, Toronto, ON, CA

²Department of Ecology and Evolutionary Biology, UMich, Ann Arbor, MI, US

[§]To whom correspondence should be addressed: hanadomoncho@gmail.com

Abstract

We investigated drought growth responses in *Medicago lupulina* using PEG to simulate drought stress. We grew *Medicago lupulina* plants inoculated with *Sinorhizobium meliloti* in Magenta boxes under randomly assigned treatments: a control, PEG applied to the bottom (PEG added to the bottom-watering container), or PEG applied from the top (PEG poured over the growth media). PEG treatments significantly reduced above-ground growth but unexpectedly increased nodulation. Our results suggest that while PEG effectively simulates drought stress on above-ground growth parameters, it may not accurately simulate drought effects on rhizobial symbiosis.

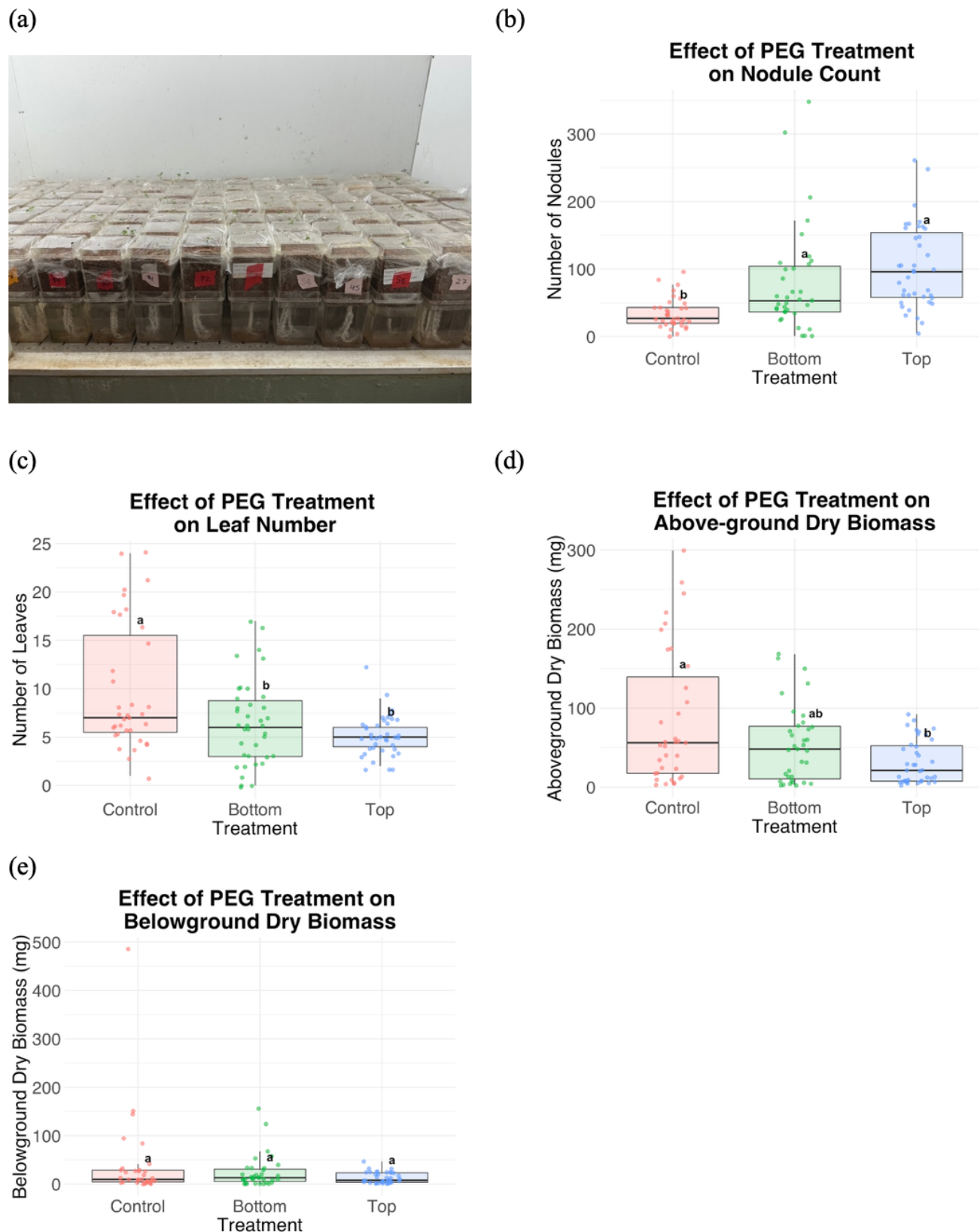


Figure 1. Experimental setup and results:

(a) Seed preparation and planting setup for *Medicago lupulina*. Individual seedlings were planted in Magenta boxes, covered with Saran Wrap and placed inside a growth chamber (25°C, 14-hour photoperiod). Effects of PEG treatments on *Medicago lupulina* growth. (b) Total nodule count, (c) number of leaves per plant across three treatments, (d) above-ground dry biomass (mg) and (e) below-ground dry biomass (mg): control, PEG applied from below (“Bottom”), and PEG applied from above (“Top”). Different letters indicate significant differences among treatments (Tukey HSD, $p < 0.05$).

Description

With rapid and unpredictable climate change, the ability of crops to withstand drought is a growing concern (Juenger and Verslues 2023; Monteleone et al. 2023; Yuan et al. 2024). Water availability is an important factor that dictates plant evolution, ecological dynamics, and physiology (Juenger and Verslues 2023), and it is vital in agriculture, where water limitation reduces yields (Begna 2020; Moss et al. 2024). Therefore, as droughts become increasingly unpredictable and severe, understanding how plants respond and adapt to water stress is crucial for understanding plant population dynamics and evolution, and improving agricultural resilience (Begna 2020; Yuan et al. 2024). Here we investigated the drought responses of *Medicago lupulina* (Black medic) using simulated drought treatments with polyethylene glycol (PEG).

Manipulative experiments that simulate drought in the greenhouse, growth chamber, or laboratory are a useful tool for studying plant physiological and evolutionary responses to drought (Verslues et al. 2006), but they can be time and labor intensive. Common approaches include applying different watering regimes (Heschel et al. 2005; Puértolas et al. 2017), using shelters or devices to divert rainfall (Hoover et al. 2018; Boyle et al. 2024), conducting dry down experiments (Puértolas et al. 2017; Juenger and Verslues 2023; Boyle et al. 2025), or using Polyethylene glycol (PEG) (Verslues et al. 2006; Osmolovskaya et al. 2018). PEG is a widely used experimental tool because of its ease of use and because it creates osmotic stress without entering plant tissues (Burnett et al. 2005; Verslues et al. 2006). However, PEG-induced drought has been shown to decrease hypocotyl elongation, germination rate, ion uptake, and gene expression compared to those under real drought conditions (Burnett et al. 2005; Verslues et al. 2006; Osmolovskaya et al. 2018; Qi et al. 2023; Kylyshbayeva et al. 2024). Accordingly, there is a need to further characterize plant responses to PEG.

Medicago lupulina is an early successional, short-lived annual legume that plays an important ecological role through its symbiotic relationship with rhizobia, which facilitates nitrogen fixation and enhances soil moisture retention (Turkington and Cavers 1979). The species has a broad global distribution across North Africa, Europe, Asia, Australia, and North America, where it grows in dry grasslands, pastures, and along roadsides (Turkington and Cavers 1979; Clark 2008), and it is widely used as a forage and fodder crop (Heuzé et al. 2018). *Medicago lupulina* can grow at elevations up to 3,600 m in the Himalayas and tolerates a wide range of environmental conditions, including mean annual temperatures between 5.7°C and 22.5°C and annual precipitation levels from 310 mm to 1,710 mm (Duke 1981; Heuzé et al. 2018), suggesting it regularly experiences variable water availability.

We tested the effects of PEG-simulated drought on *Medicago lupulina* and its mutualistic interactions using three experimental treatments: a control, where plants received only water; a treatment where 150g L⁻¹ 8000-PEG solution was poured over the growth media; and a treatment where the same solution was added to the bottom watering container of the magenta box (Figure 1a, see *Methods*; Burnett et al. 2005; Piwowarczyk et al. 2014; Castañeda and González 2021). We germinated seeds of *Medicago lupulina* and inoculated plants with mutualistic *Sinorhizobium meliloti* rhizobacteria. We applied nitrogen-free Jensen's fertilizer (Boyle et al. 2021) after 26 days and ended the experiment after 60 days.

Our results show that PEG treatment significantly reduced above-ground performance, specifically the number of leaves and above-ground biomass (Figure 1c and 1d). Control plants had significantly larger above-ground biomass than those top-watered with PEG solution, while those bottom-watered with PEG were not significantly different from controls or top-watering PEG-treated plants (Figure 1e). These data suggest that PEG can effectively simulate drought stress for above-ground growth metrics. However, we also detected a significant increase in nodule number in response to PEG application (Figure 1b). The usual effect of drought on nodulation is nodule senescence and decreased nodulation (e.g., Arrese-Igor et al. 2011; Iqbal et al. 2022; Istanbul et al. 2022; Melo et al. 2025; Ruiz-Lozano et al. 2001; for a phylogenetic meta-analysis, including species of *Medicago*, see Iqbal et al. 2022). Mhadhbi et al. (2009) showed decreased nodule number in *Medicago truncatula* in response to osmotic stress from mannitol treatments, while Dhanushkodi et al. (2018) showed that reduced soil water content in the *Medicago truncatula* led to an ~50% reduction in mean nodule number relative to well-watered controls. Our findings of significantly increased nodulation in both PEG treatments relative to the control are thus in contrast with the prevailing reports in the literature for legumes, phylogenetic meta-analyses, and experimental results using simulated and actual drought in congeneric *Medicago* species. We originally hypothesized that plants might compensate for simulated drought by investing more in below-ground growth, which could explain higher nodule counts (Koziol et al. 2012; Sofi et al. 2018; Lumactud et al. 2023). However, our measurements of dry belowground biomass showed no significant differences among treatments (Figure 1e), which does not support this explanation. Multiple alternative hypotheses could explain the finding of increased nodulation in response to PEG. It could be that PEG solution, a persistent liquid that the plants could not absorb, was favorable for rhizobia mobility and/or infection, resulting in increased nodulation. Alternatively, PEG may affect plant production of flavonoids (Sarmadi et al. 2019), a key step in the nodulation process (Subramanian et al. 2007), or the dispersal of flavonoids away from the roots, both of which could have led to increased nodulation. In addition, osmotic stress has been shown to increase the expression of Nod genes in other species of *Sinorhizobium* (Fuentes-Romero et al. 2023), which if it occurred in this experiment, could also have led to increased nodulation. Regardless of the mechanism, our results suggest caution in the use of PEG to simulate the effect of drought on plant-rhizobia mutualisms.

Methods

Seed Preparation

We scarified a total of 150 *M. lupulina* seeds, sterilized them in 100% ethanol, and imbibed them in distilled water. The seeds were placed on 1% agar plates, wrapped in aluminium foil, and stored at 4 °C for eight days (Simonsen and Stinchcombe 2014; Boyle et al. 2021). We then moved them to room temperature (~24 °C) for 16 hours to allow radicle emergence (Simonsen and Stinchcombe 2014; Boyle et al. 2021). Germinated seeds were planted individually in Magenta boxes. Each box consisted of a top chamber filled with sterilized Turface and a cotton string wick extending through a hole at the bottom to draw water from a lower chamber. The setup was covered with Saran Wrap in an attempt to minimize sources of dehydration other than those caused by PEG; when plants grew tall enough, we poked a small hole in the Saran Wrap and gently pulled stems through. We placed magenta boxes in a growth chamber with a 14-hour photoperiod at 25°C; we cultured the bacterial strain *Sinorhizobium meliloti* WSM1022 following Simonsen and Stinchcombe (2014), and applied 1 mL of inoculum the day after planting (Figure 1d). After 26 days of growth, we applied 1 mL of nitrogen-free Jensen's fertilizer.

PEG Treatments

We prepared 150 g L⁻¹ polyethylene glycol (PEG 8000) solution to simulate drought. Plants were randomly assigned to one of three treatments: a control with 150 mL of sterilized water added to the bottom chamber, a "Bottom" treatment where 150 mL of PEG solution was added to the bottom chamber, and a "Top" treatment where 150 mL of PEG solution was applied to the soil surface. We included both the top and bottom treatments because it was unclear how the PEG solution would interact with the wicks used in the Magenta boxes; the top treatment ensured that plants and their roots would be exposed to PEG in the growth media. We applied the treatments 30 days after planting, with no further water or PEG solution supplied for the remainder of the experiment.

Measurements

We counted the number of true leaves 40 days after they were planted. On day 50, we destructively harvested four plants to visually confirm nodule formation; these samples were excluded from further analyses. Because of these destructive samples, and some germinants failing to establish, final sample sizes were 43 controls, 44 in the bottom PEG treatment, and 43 in the top PEG treatment. We took measurements 60 days after planting, including true leaf and nodule counts, and dried above-ground dry biomass. We initially stored roots in a lab fridge, then oven-dried and weighed them to obtain below-ground dry biomass.

Statistical Analysis

We conducted the analysis in R (v4.5.1; R Core Team 2025) using base R functions and the *tidyverse*, *ggplot2*, and *multcompView* packages for data processing and visualization (Graves et al. 2024; Wickham 2019; Wickham et al. 2019). For below-ground biomass, a single data point was 8.8 standard deviations from the mean and was removed as an outlier. We used a one-way analysis of variance (ANOVA) function, *ao**v*(), to evaluate the effect of the different PEG treatments on leaf number, nodule number, and below- and above-ground dry biomass. When ANOVA results indicated significant treatment effects ($p < 0.05$), we conducted pairwise comparisons among treatments using Tukey's Honestly Significant Difference (HSD) tests. Significance groupings were extracted using the *multcompLetters4*() function and we created a box plot with overlaid jittered data points to display individual variation. Raw data and R code for this work are archived at Zenodo (Cho et al. 2026).

Acknowledgements: We thank the EEB horticultural staff for advice and support.

References

- Arrese-Igor, Cesar, Esther M. González, Daniel Marino, Rubén Ladrera, Estíbaliz Larrainzar, and Erena Gil-Quintana. Physiological responses of legume nodules to drought. *Plant stress* 5, no. 1 (2011): 24-31.
- Begna T. 2020. Effects of Drought Stress on Crop Production and Productivity. *International Journal of Research Studies in Agricultural Sciences*. 6(9). <https://doi.org/10.20431/2454-6224.0609005>
- Boyle JA et al. 2024. Resistance and resilience of soil microbiomes under climate change. *Ecosphere*. 15(12):e70077. <https://doi.org/10.1002/ecs2.70077>
- Boyle JA et al. 2025. Mutualism mediates legume response to microbial climate legacies. *Ecology and Evolution*. 15(10):e72271. <https://doi.org/10.1002/ece3.72271>
- Boyle JA, Simonsen AK, Frederickson ME, Stinchcombe JR. 2021. Priority effects alter interaction outcomes in a legume–rhizobium mutualism. *Proc R Soc B*. 288(1946):20202753. <https://doi.org/10.1098/rspb.2020.2753>
- Burnett S, Thomas P, van Iersel M. 2005. Postgermination drenches with PEG-8000 reduce growth of *Salvia* and *Marigolds*. *HortScience*. 40(3):675–679. <https://doi.org/10.21273/HORTSCI.40.3.675>

- Castañeda V, González EM. 2021. Strategies to apply water-deficit stress: Similarities and disparities at the whole plant metabolism level in *Medicago truncatula*. *IJMS*. 22(6):2813. <https://doi.org/10.3390/ijms22062813>
- Cho H, Stinchcombe JR, Glasgow E, Mukund V, Boyle JA. 2026. Dataset and R code for simulated drought with polyethylene glycol on *Medicago lupulina* [dataset]. Zenodo. <https://doi.org/10.5281/zenodo.18523987> DOI: <https://doi.org/10.5281/zenodo.18523987>
- Clark, A. (Ed.). (2008). Managing cover crops profitably. 3rd ed. Sustainable Agriculture Research and Education (SARE) program. University of Maryland, College Park, MD, USA
- Dhanushkodi, R., Matthew, C., McManus, M. T., & Dijkwel, P. P. (2018). Drought-induced senescence of *Medicago truncatula* nodules involves serpin and ferritin to control proteolytic activity and iron levels. *The New Phytologist*, 220(1), 196–208.
- Duke JA. 1981. Handbook of LEGUMES of World Economic Importance. 1st ed. Plenum Press, New York, USA. <https://doi.org/10.1007/978-1-4684-8151-8>
- Fuentes-Romero F et al. 2023. Non-Ionic Osmotic Stress Induces the Biosynthesis of Nodulation Factors and Affects Other Symbiotic Traits in *Sinorhizobium fredii* HH103. *Biology*. 12(2):148. <https://doi.org/10.3390/biology12020148>
- Graves S, Piepho H-P, Selzer L, Dorai-Raj S. 2024. multcompView: Visualizations of Paired Comparisons. Version 0.1-10. <https://doi.org/10.32614/CRAN.package.multcompView>
- Heschel MS, Hausmann N, Schmitt J. 2005. Testing for stress-dependent inbreeding depression in *Impatiens capensis* (Balsaminaceae). *American J of Botany*. 92(8):1322–1329. <https://doi.org/10.3732/ajb.92.8.1322>
- Heuzé V et al. 2018. Black medic (*Medicago lupulina*). Feedipedia, a programme by INRAE, CIRAD, AFZ and FAO. [accessed 2025 Oct 10]. <https://www.feedipedia.org/node/277>
- Hoover DL, Wilcox KR, Young KE. 2018. Experimental droughts with rainout shelters: a methodological review. *Ecosphere*. 9(1):e02088. <https://doi.org/10.1002/ecs2.2088>
- Iqbal, N., Sadras, V. O., Denison, R. F., Zhou, Y., & Denton, M. D. (2022). Clade-dependent effects of drought on nitrogen fixation and its components – Number, size, and activity of nodules in legumes. *Field Crops Research*, 284(108586), 108586.
- Istanbuli T et al. 2022. The interaction between drought stress and nodule formation under multiple environments in chickpea Gahlaut V, editor. *PLoS ONE*. 17(10):e0276732. <https://doi.org/10.1371/journal.pone.0276732>
- Juenger TE, Verslues PE. 2023. Time for a drought experiment: Do you know your plants' water status? *The Plant Cell*. 35(1):10–23. <https://doi.org/10.1093/plcell/koac324>
- Kylyshbayeva G, Bishimbayeva N, Jatayev S, Eliby S, & Shavrukov Y. 2025. Polyethylene Glycol (PEG) application triggers plant dehydration but does not accurately simulate drought. *Plants*. 14(1): 92. <https://doi.org/10.3390/plants14010092>
- Kozioł L, Rieseberg LH, Kane N, Bever JD. 2012. Reduced drought tolerance during domestication and the evolution of weediness results from tolerance-growth trade-offs. *Evolution*. 66(12):3803–3814. <https://doi.org/10.1111/j.1558-5646.2012.01718.x>
- Lumactud RA et al. 2023. The effect of drought stress on nodulation, plant growth, and nitrogen fixation in soybean during early plant growth. *J Agronomy Crop Science*. 209(3):345–354. <https://doi.org/10.1111/jac.12627>
- Melo AAR, Araújo MA, Mendes NAC, Reis AR. 2025. Drought stress disrupts biological nitrogen fixation and starch accumulation compromising growth and yield of cowpea plants. *Plant Physiology and Biochemistry*. 224:109931. <https://doi.org/10.1016/j.plaphy.2025.109931>
- Mhadhbi, H., Fotopoulos, V., Djebali, N., Polidoros, A. N., & Aouani, M. E. (2009). Behaviours of *Medicago truncatula*-*Sinorhizobium meliloti* Symbioses under osmotic stress in relation with the symbiotic partner input: Effects on nodule functioning and protection. *Journal of Agronomy and Crop Science*, 195(3), 225–231.
- Monteleone B, Borzì I, Bonaccorso B, Martina M. 2023. Quantifying crop vulnerability to weather-related extreme events and climate change through vulnerability curves. *Nat Hazards*. 116(3):2761–2796. <https://doi.org/10.1007/s11069-022-05791-0>
- Moss WE et al. 2024. Drought as an emergent driver of ecological transformation in the twenty-first century. *BioScience*. 74(8):524–538. <https://doi.org/10.1093/biosci/biae050>
- Osmolovskaya N et al. 2018. Methodology of drought stress research: Experimental setup and physiological characterization. *IJMS*. 19(12):4089. <https://doi.org/10.3390/ijms19124089>

- Piwowarczyk B, Kamińska I, Rybiński W. 2014. Influence of PEG generated osmotic stress on shoot regeneration and some biochemical parameters in *Lathyrus* culture. Czech J. Genet. Plant Breed., 50, 2014 (2): 77–83 <https://doi.org/10.13140/2.1.4330.3040>
- Puértolas J, Larsen EK, Davies WJ, Dodd IC. 2017. Applying ‘drought’ to potted plants by maintaining suboptimal soil moisture improves plant water relations. Journal of Experimental Botany. 68(9):2413–2424. <https://doi.org/10.1093/jxb/erx116>
- Qi Y et al. 2023. Effects of drought stress induced by hypertonic Polyethylene Glycol (PEG-6000) on *Passiflora edulis* Sims physiological properties. Plants. 12(12):2296. <https://doi.org/10.3390/plants12122296>
- R Core Team (2025). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. <https://www.R-project.org/>
- Ruis-Lozano JM, Collados C, Barea JM, Azcón R. 2001. Arbuscular mycorrhizal symbiosis can alleviate drought – induced nodule senescence in soybean plants. New Phytologist. 151(2):493–502. <https://doi.org/10.1046/j.0028-646x.2001.00196.x>
- Sarmadi M et al. 2019. Improved effects of polyethylene glycol on the growth, antioxidative enzymes activity and taxanes production in a *Taxus baccata* L. callus culture. Plant Cell Tiss Organ Cult. 137(2):319–328. <https://doi.org/10.1007/s11240-019-01573-y>
- Simonsen AK, Stinchcombe JR. 2014. Standing genetic variation in host preference for mutualist microbial symbionts. Proceedings. Biological sciences, 281(1797), 20142036. <https://doi.org/10.1098/rspb.2014.2036>
- Sofi PA, Djanaguiraman M, Siddique KHM, Prasad PVV. 2018. Reproductive fitness in common bean (*Phaseolus vulgaris* L.) under drought stress is associated with root length and volume. Ind J Plant Physiol. 23(4):796–809. <https://doi.org/10.1007/s40502-018-0429-x>
- Subramanian S, Stacey G, Yu O. 2007. Distinct, crucial roles of flavonoids during legume nodulation. Trends in Plant Science. 12(7):282–285. <https://doi.org/10.1016/j.tplants.2007.06.006>
- Turkington R, Cavers PB. 1979. The biology of Canadian weeds.: 33. *Medicago lupulina* L. Can J Plant Sci. 59(1):99–110. <https://doi.org/10.4141/cjps79-015>
- Uzun F, Aydin I. 2004. Improving germination rate of *Medicago* and *Trifolium* species. Asian J of Plant Sciences. 3(6):714–717. <https://doi.org/10.3923/ajps.2004.714.717>
- Verslues PE et al. 2006. Methods and concepts in quantifying resistance to drought, salt and freezing, abiotic stresses that affect plant water status. The Plant Journal. 45(4):523–539. <https://doi.org/10.1111/j.1365-3113.2005.02593.x>
- Wickham H. ggplot2: Elegant Graphics for Data Analysis. Springer-Verlag New York, 2016.
- Wickham H et al. 2019. Welcome to the Tidyverse. JOSS. 4(43):1686. <https://doi.org/10.21105/joss.01686>
- Wickham H et al. 2019. Welcome to the tidyverse. Journal of Open Source Software. 4(43). 1686. <https://doi.org/10.21105/joss.01686>.
- Yuan X et al. 2024. Impacts of global climate change on agricultural production: A comprehensive review. Agronomy. 14(7):1360. <https://doi.org/10.3390/agronomy14071360>

Funding: University of Toronto’s Centre for Global Change Science (HC), NSERC Discovery Grants (JRS), and NSERC doctoral post-graduate fellowships (JAB).

Conflicts of Interest: The authors declare that there are no conflicts of interest present.

Author Contributions: Hana Cho: conceptualization, data curation, formal analysis, investigation, methodology, visualization, writing - original draft, writing - review editing. Emily Glasgow: conceptualization, data curation, investigation, methodology, writing - review editing. Valmic Mukund: methodology, writing - review editing, resources. Julia A. Boyle: conceptualization, methodology, writing - review editing. John R. Stinchcombe: conceptualization, formal analysis, funding acquisition, methodology, project administration, supervision, resources, writing - original draft, writing - review editing.

Reviewed By: Anonymous

History: Received December 20, 2025 **Revision Received** February 8, 2026 **Accepted** February 13, 2026 **Published Online** February 17, 2026 **Indexed** March 3, 2026

Copyright: © 2026 by the authors. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International (CC BY 4.0) License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

2/17/2026 - Open Access

Citation: Cho H, Glasgow E, Mukund V, Boyle JA, Stinchcombe JR. 2026. Simulated drought with Polyethylene-Glycol (PEG) decreases above-ground performance and increases nodulation in the legume *Medicago lupulina*. microPublication Biology. [10.17912/micropub.biology.001997](https://doi.org/10.17912/micropub.biology.001997)