

Transcripts from the *src-1(cj293)* mutant can encode a SRC-1 molecule lacking the SH2 domain in *Caenorhabditis elegans*

Snehal S Mahadik¹, Erik A. Lundquist^{1§}

¹Molecular Biosciences, University of Kansas, Lawrence, Kansas, United States

[§]To whom correspondence should be addressed: erikl@ku.edu

Abstract

Previous studies suggest that the [src-1\(cj293\)](#) mutation is an activated [src-1](#) allele in *C. elegans* with the potential to encode a molecule lacking the SH2 domain. [src-1\(cj293\)](#) is a deletion with breakpoints in introns 3 and 5, deleting exons 4 and 5, which encode the SH2 domain. If exon 3 is spliced to exon 6, the reading frame is maintained. Here, RNA seq of [src-1\(cj293\)](#) mutants showed that the exon 3 to exon 6 splice does not occur in [src-1\(+\)](#) but is robustly present in [src-1\(cj293\)](#). Thus, [src-1\(cj293\)](#) produces a transcript that can encode a [SRC-1](#) molecular lacking the SH2 domain, which leads to overactive [SRC-1](#) in growth cones of VD neurons during their outgrowth (*i.e.* [src-1\(cj293\)](#) might be a constitutively-active mutation).

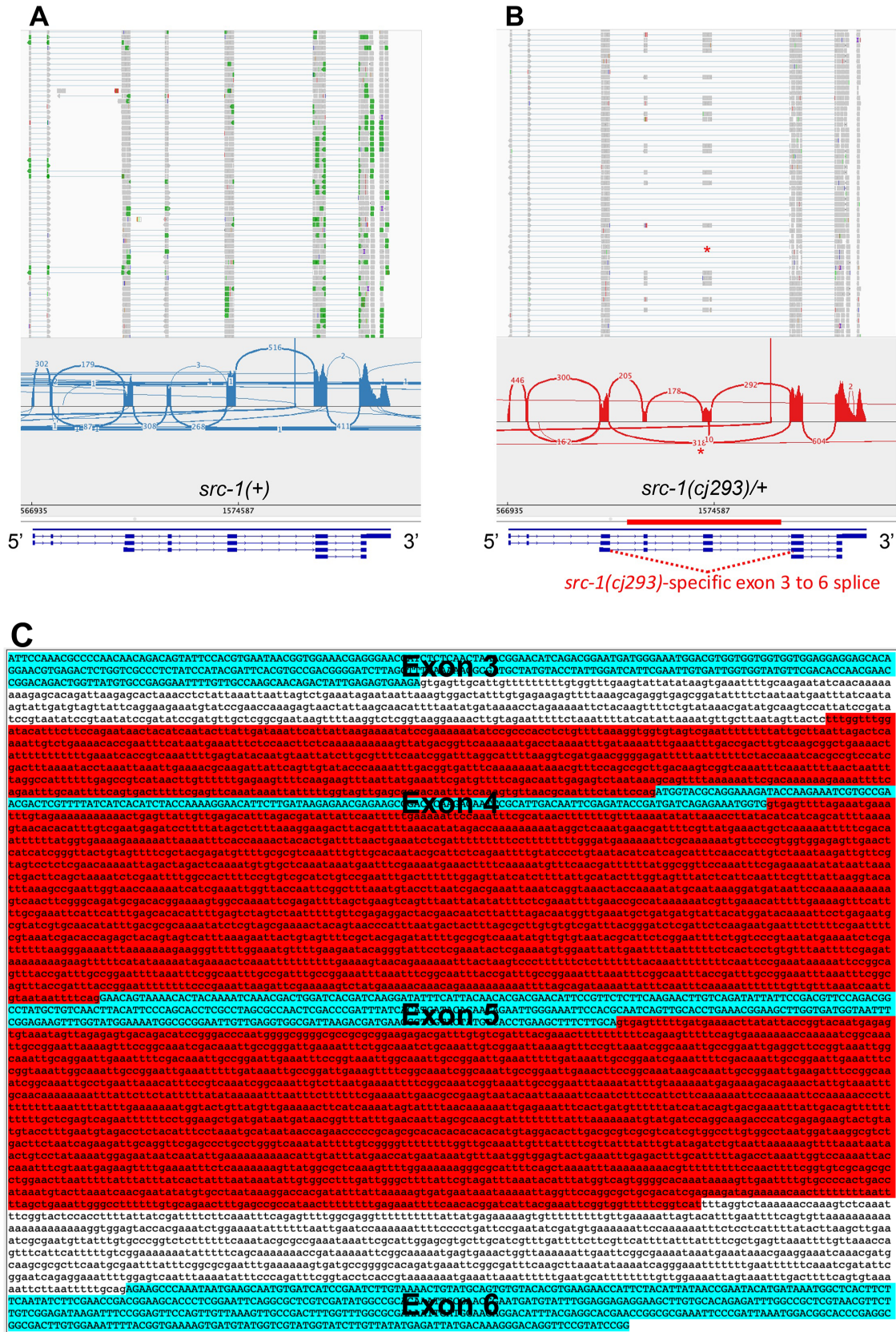


Figure 1. Transcripts from the *src-1* locus in *src-1*(+) and *src-1*(*cj293*)/+:

Alignments of RNA seq to the *src-1* locus in A) *src-1(+)* and B) *src-1(cj293)*. At the top are representative alignments from the Integrated Genome Viewer. The asterisk in *src-1(cj293)/+* indicates a group of exon 3 to exon 6 splices, which did not occur in *src-1(+)*. Shown below the alignments are Sashimi plots of splicing at the *src-1* locus, generated in the Integrated Genome Viewer. The 318 exon 3 to exon 6 splices are indicated with an asterisk. The structure of the *src-1* locus is shown at the bottom. The extent of the *src-1(cj293)* deletion is indicated with a red bar. 5' to 3' orientation of the locus is indicated. The *src-1(cj293)*-specific exon 3 to 6 splice is indicated with a dashed red line below the gene model. C) The sequence of the *src-1* locus from exon 3 to exon 6 (bases 1,570,337 to 1,577,868 on LGI). The region deleted in *src-1(cj293)* is red, and the exons are cyan. Exons 4 and 5 are removed by the *src-1(cj293)* deletion.

Description

The **SRC-1**/Src tyrosine kinase in *Caenorhabditis elegans* is required for embryonic development, cell migration, and axon guidance (Bei *et al.* 2002; Itoh *et al.* 2005; Lee *et al.* 2005; Sugioka and Sawa 2010; Masuda *et al.* 2012; Zhu *et al.* 2020; Mahadik *et al.* 2024). In developing VD axon growth cones, **SRC-1** acts with the **UNC-6**/Netrin receptor **UNC-5** to inhibit growth cone protrusion (Mahadik *et al.* 2024). A precise deletion mutant of *src-1* had VD growth cones that displayed excessive protrusion resulting in axon guidance defects, similar to *unc-5* loss-of-function (Mahadik *et al.* 2024). The *src-1(cj293)* in-frame deletion removes exons that encode the SH2 domain and an N-terminal portion of the kinase domain (Mahadik *et al.* 2024). *src-1(cj293)* is predicted to encode a molecule lacking the SH2 domain and part of the kinase domain (Mahadik *et al.* 2024). Similar to the *src-1(lq185)* precise deletion allele, *src-1(cj293)* mutants display defects in embryonic development (Bei *et al.* 2002; Mahadik *et al.* 2024). However, *src-1(cj293)* mutants displayed VD growth cones with reduced protrusion compared to *wild-type*, similar to *src-1(+)* overexpression (Mahadik *et al.* 2024). This suggests that *src-1(cj293)* might encode a constitutively-active **SRC-1** molecule.

The breakpoints of the *src-1(cj293)* mutation are in introns 3 and 5, removing exons 4 and 5 (Figure 1A and B). If exon 3 is spliced to exon 6 (3-6), the reading frame is maintained, resulting in coding potential for a molecule lacking the SH2 domain and part of the kinase domain (Mahadik *et al.* 2024). RNA seq was conducted in heterozygous *src-1(cj293)/+* animals. Reads were aligned to the *C. elegans* genome, and Sashimi plots were generated to illustrate splicing events. In animals with a *wild-type src-1(+)* gene, the 3-6 splice did not occur (Figure 1A). However, in *src-1(cj293)/+* animals, the 3-6 splice was common (318 times) (Figure 1B). The 3-6 splice products produced in *src-1(cj293)* have the potential to produce a **SRC-1** molecule missing the SH2 domain and part of the kinase domain. The catalytic residue of the kinase is not in the deleted region. Kinase function is likely active in *src-1(cj293)*, as mutation of the catalytic residue (D381A) in *src-1(syb7248)* resulted in a dominant phenotype resembling *src-1* precise deletion, with excessively-protrusive growth cones VD growth cones (Mahadik *et al.* 2024).

Autoinhibition of Src kinase activity is mediated by the SH2 domain, which binds to phosphorylated tyrosine 527, resulting in a closed, inactive conformation (reviewed in (Wagner *et al.* 2013)). This tyrosine is conserved in *C. elegans SRC-1* (tyrosine 531), thus it might also be subject to autoinhibition by the SH2 domain. A **SRC-1** molecule missing the SH2 domain may result in overactive kinase activity, consistent with the *src-1(cj293)* overactive phenotype in the growth cone. It is also possible that *src-1(cj293)* overactivity is due to other interactions that require the SH2 domain and/or the N-terminal portion of the kinase domain. *src-1(cj293)* was not dominant for axon guidance defects, which would be expected of an activated molecule. Possibly, one copy of the activated allele is not sufficient to produce a phenotype in heterozygotes with *src-1(+)*. In any case, these results are consistent with *src-1(cj293)* producing an activated **SRC-1** molecule lacking the SH2 domain and N-terminal portion of the kinase domain, which phenotypically, results in **SRC-1** overactivity.

Methods

RNA was isolated from mixed-stage animals as previously described (Tamayo *et al.* 2013; Paolillo *et al.* 2024). Poly-A selection and RNA seq library construction was conducted using the NEBnext stranded RNA seq kit. RNA seq libraries were made using the NEBNext stranded mRNA library kit. Sequencing was conducted on a Nextseq 2000 instrument with 150-bp paired-end sequencing. FASTQ files were processed using fastp (0.23.2) (Chen *et al.* 2018). Reads were aligned to the *C. elegans* reference genome [release WBcel235, version WBPS14 (WS271)] using HISAT2 (version 2.2.1) (Kim *et al.* 2015). BAM files from HISAT2 alignment were analyzed in the Integrated Genome Viewer (Robinson *et al.* 2011; Thorvaldsdottir *et al.* 2012), including Sashimi plots (Katz *et al.* 2010; Katz *et al.* 2015). Wormbase was used for *C. elegans* informatics (Sternberg *et al.*, 2024)

Reagents

Raw FASTQ reads for *src-1(+)* and *src-1(cj293)* were deposited in the Sequence Read Archive ([PRJNA1093133](#) and [PRJNA1219192](#), respectively). The *src-1(+)* strain was LE5443 (*unc-6(lq154) X; juIs76 II*). The balanced *src-1(cj293)/+* strain was [HR1275](#) (*src-1(cj293) dpy-5(e61)/hT2 I; +/hT2 III*).

Acknowledgements: The authors thank members of the Lundquist lab for discussions, and Wormbase for *C. elegans* informatics. Some strains were provided by the CGC, which is funded by NIH Office of Research Infrastructure Programs (P40 OD010440).

References

Bei Y, Hogan J, Berkowitz LA, Soto M, Rocheleau CE, Pang KM, Collins J, Mello CC. 2002. SRC-1 and Wnt Signaling Act Together to Specify Endoderm and to Control Cleavage Orientation in Early *C. elegans* Embryos. *Developmental Cell* 3: 113-125. DOI: [10.1016/s1534-5807\(02\)00185-5](#)

Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor. *Bioinformatics* 34: i884-i890. DOI: [10.1093/bioinformatics/bty560](#)

Itoh B, Hirose T, Takata N, Nishiwaki K, Koga M, Ohshima Y, Okada M. 2005. SRC-1, a non-receptor type of protein tyrosine kinase, controls the direction of cell and growth cone migration in *C. elegans*. *Development* 132: 5161-5172. DOI: [dev.02103 \[pii\] 10.1242/dev.02103](#)

Katz Y, Wang ET, Airoidi EM, Burge CB. 2010. Analysis and design of RNA sequencing experiments for identifying isoform regulation. *Nature Methods* 7: 1009-1015. DOI: [10.1038/nmeth.1528](#)

Katz Y, Wang ET, Silterra J, Schwartz S, Wong B, Thorvaldsdóttir H, et al., Burge. 2015. Quantitative visualization of alternative exon expression from RNA-seq data. *Bioinformatics* 31: 2400-2402. DOI: [10.1093/bioinformatics/btv034](#)

Kim D, Langmead B, Salzberg SL. 2015. HISAT: a fast spliced aligner with low memory requirements. *Nature Methods* 12: 357-360. DOI: [10.1038/nmeth.3317](#)

Lee J, Li W, Guan KL. 2005. SRC-1 Mediates UNC-5 Signaling in *Caenorhabditis elegans*. *Molecular and Cellular Biology* 25: 6485-6495. DOI: [10.1128/MCB.25.15.6485-6495.2005](#)

Mahadik SS, Burt EK, Lundquist EA. 2024. SRC-1 controls growth cone polarity and protrusion with the UNC-6/Netrin receptor UNC-5 in *Caenorhabditis elegans*. *PLOS ONE* 19: e0295701. DOI: [10.1371/journal.pone.0295701](#)

Masuda H, Nakamura K, Takata N, Itoh B, Hirose T, Moribe H, Mekada E, Okada M. 2012. MIG-13 controls anteroposterior cell migration by interacting with UNC-71/ADM-1 and SRC-1 in *Caenorhabditis elegans*. *FEBS Letters* 586: 740-746. DOI: [10.1016/j.febslet.2012.01.031](#)

Paolillo VK, Ochs ME, Lundquist EA. 2024. MAB-5/Hox regulates the Q neuroblast transcriptome, including *cwn-1/Wnt*, to mediate posterior migration in *Caenorhabditis elegans*. *GENETICS* 227: 10.1093/genetics/iyae045. DOI: [10.1093/genetics/iyae045](#)

Robinson JT, Thorvaldsdóttir H, Winckler W, Guttman M, Lander ES, Getz G, Mesirov JP. 2011. Integrative genomics viewer. *Nature Biotechnology* 29: 24-26. DOI: [10.1038/nbt.1754](#)

Sternberg PW, Van Auken K, Wang Q, Wright A, Yook K, Zarowiecki M, et al., Stein. 2024. WormBase 2024: status and transitioning to Alliance infrastructure. *GENETICS* 227: 10.1093/genetics/iyae050. DOI: [10.1093/genetics/iyae050](#)

Sugioka K, Sawa H. 2010. Regulation of asymmetric positioning of nuclei by Wnt and Src signaling and its roles in POP-1/TCF nuclear asymmetry in *Caenorhabditis elegans*. *Genes to Cells* 15: 397-407. DOI: [10.1111/j.1365-2443.2010.01388.x](#)

Tamayo JV, Gujar M, Macdonald SJ, Lundquist EA. 2013. Functional transcriptomic analysis of the role of MAB-5/Hox in Q neuroblast migration in *Caenorhabditis elegans*. *BMC Genomics* 14: 10.1186/1471-2164-14-304. DOI: [10.1186/1471-2164-14-304](#)

Thorvaldsdottir H, Robinson JT, Mesirov JP. 2012. Integrative Genomics Viewer (IGV): high-performance genomics data visualization and exploration. *Briefings in Bioinformatics* 14: 178-192. DOI: [10.1093/bib/bbs017](#)

Wagner MJ, Stacey MM, Liu BA, Pawson T. 2013. Molecular Mechanisms of SH2- and PTB-Domain-Containing Proteins in Receptor Tyrosine Kinase Signaling. *Cold Spring Harbor Perspectives in Biology* 5: a008987-a008987. DOI: [10.1101/cshperspect.a008987](#)

Zhu Z, Chai Y, Hu H, Li W, Li WJ, Dong MQ, et al., Ou. 2020. Spatial confinement of receptor activity by tyrosine phosphatase during directional cell migration. *Proceedings of the National Academy of Sciences* 117: 14270-14279. DOI:

3/12/2025 - Open Access

[10.1073/pnas.2003019117](https://doi.org/10.1073/pnas.2003019117)

Funding: RNA seq was conducted in the University of Kansas Genome Sequencing Core Laboratory funded by the NIH *Center for Molecular Analysis of Disease Pathways* (P30GM145499). The NIH *Kansas Infrastructure Network of Biomedical Research Excellence* provided infrastructure support (P20GM103418). Supported by National Institute of Neurological Disorders and Stroke (United States) R03NS114554 to Erik A. Lundquist. ,Supported by National Institute of Neurological Disorders and Stroke (United States) R01NS115467 to Erik A. Lundquist. ,Supported by National Institute of General Medical Sciences (United States) P30GM145499 to Erik A. Lundquist.

Author Contributions: Snehal S Mahadik: writing - review editing, conceptualization. Erik A. Lundquist: conceptualization, data curation, funding acquisition, investigation, methodology, formal analysis, project administration, writing - original draft, writing - review editing.

Reviewed By: Anonymous

Nomenclature Validated By: Stavros Diamantakis, Anonymous

WormBase Paper ID: WBPaper00067831

History: **Received** February 10, 2025 **Revision Received** February 20, 2025 **Accepted** March 4, 2025 **Published Online** March 12, 2025 **Indexed** March 26, 2025

Copyright: © 2025 by the authors. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International (CC BY 4.0) License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Citation: Mahadik SS, Lundquist EA. 2025. Transcripts from the *src-1(cj293)* mutant can encode a SRC-1 molecule lacking the SH2 domain in *Caenorhabditis elegans*. microPublication Biology. [10.17912/micropub.biology.001537](https://doi.org/10.17912/micropub.biology.001537)