# The *Caenorhabditis elegans* Dispatched ortholog, CHE-14, is dispensable for apical secretion of the Hedgehog-related proteins GRL-2 and WRT-10

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# Abstract

<u>*C. elegans*</u> nematodes possess expanded families of Hedgehog related (Hh-r) and Patched/Dispatched-related (PTR) proteins but their functional relationship remains unclear. Here we investigated whether <u>CHE-14</u>, the closest <u>*C. elegans*</u> ortholog for the Hedgehog transporter Dispatched, was necessary for the secretion of two tagged Hh-r proteins: <u>WRT-10</u> and <u>GRL-2</u>. We report that <u>CHE-14</u> is dispensable for the apical localization of <u>GRL-2</u> and <u>WRT-10</u>. We also show that animals lacking <u>CHE-14</u> and another redundant PTR protein <u>DAF-6</u> also secrete <u>WRT-10</u>, suggesting neither are required for secretion of these specific Hh-r proteins.

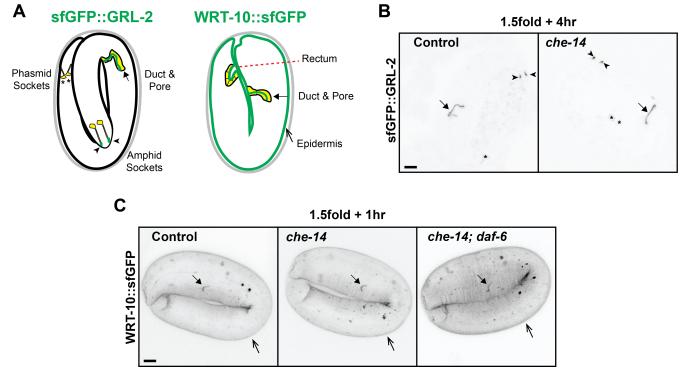


Figure 1. Hh-r proteins GRL-2 and WRT-10 localize normally in *che-14* mutant nematodes:

A) Schematic diagrams of Hh-r fusion protein localization (green) in embryos. The cytoplasm of cells of interest is indicated in yellow. sfGFP::GRL-2 localizes to the lumens of specific unicellular tubes, the amphid glia socket cells and the excretory duct and pore. sfGFP::GRL-2 is observed cytoplasmically in the phasmid socket glia, and localizes to this cell's lumen later in development (Serra et al., 2024). WRT-10::sfGFP localizes to the epidermal surface, the rectum, and the excretory duct and pore lumens. Stages and symbols match those in the micrographs below. B) sfGFP::GRL-2 localizes to the cuticle of the excretory duct (arrow) and amphid socket lumens (arrowhead) in both *che-14* and control animals. Asterisks mark the cytoplasmic expression in phasmid socket cells. C) WRT-10::sfGFP localizes to the embryonic sheath matrix lining the epidermis (open arrow) and excretory duct and pore (closed arrow) in *che-14*, *che-14*; *daf-6* double mutants, and control animals. Images shown are max projections of full-embryo z-stacks. In all panels, scale bars= 5 µm. At least n=10 animals were observed for each genotype. Embryos were obtained from homozygous mutant mothers of the indicated genotype, except in double mutant animals the mothers contained a rescue transgene (*csEx946*). Additional characterization of these fusion proteins in wild-type animals can be found in (Serra et al., 2024).



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## Description

Despite lacking clear orthologs for the Hedgehog signaling ligand and the majority of its canonical pathway partners, *Caenorhabditis elegans* nematodes possess 26 Patched/Dispatched-related (PTR) proteins (the canonical Hedgehog receptor and transporter, respectively) and over twice as many small proteins considered to be evolutionarily Hedgehog-related (Hh-r) (Bürglin and Kuwabara 2006; Bürglin 2008; Hao et al., 2006; Zugasti et al., 2005; Aspöck et al., 1999). We recently designed endogenously tagged sfGFP fusion proteins for two Hh-r proteins, <u>GRL-2</u> and <u>WRT-10</u>, and found that they were both tissue and substructure specific apical extracellular matrix (aECM) components (Serra et al., 2024). <u>GRL-2</u>, which belongs to the Groundhog-like (GRL) family, is a component of specific tube cuticles, while <u>WRT-10</u>, which belongs to the Warthog (WRT) family, is a component of the transient precuticle or "sheath" matrix that precedes the cuticle in developing worms (Figure 1A). Other Hh-r proteins, such as <u>GRL-7</u> and <u>GRL-18</u>, have also been localized to specific aECMs (Chiyoda et al., 2021; Fung et al., 2023). Many PTR proteins also affect the aECM (reviewed Sundaram and Pujol 2024), but the specific relationships between Hh-r and PTR proteins remain unclear.

In canonical Hedgehog signaling, Dispatched is a necessary mediator of Hedgehog trafficking and release (reviewed in Zhang and Beachy 2023). Among <u>*C. elegans*</u> proteins, the PTR <u>CHE-14</u> shares the most sequence homology with Dispatched (Michaux et al., 2000; Bürglin and Kuwabara 2006). Furthermore, <u>CHE-14</u> has previously reported roles in apical secretion to promote proper aECM organization in the epidermis, excretory system tubes, and amphid glia (Michaux et al., 2000; Perens and Shaham 2005). Therefore, we hypothesized that <u>CHE-14</u> might promote the apical secretion of Hh-r proteins from these tissues.

Here we show that, counter to our hypothesis, both <u>GRL-2</u> and <u>WRT-10</u> sfGFP fusion proteins are still properly secreted and localize to matrix in <u>*che-14(ok193)*</u> null mutant animals. In both wild-type and <u>*che-14*</u> mutant embryos, <u>GRL-2</u> localizes to the cuticle matrix lining the amphid socket and the excretory duct and pore lumens (Figure 1B). <u>WRT-10</u> also localizes to the apical matrix of the epidermis, the rectum, and the excretory duct and pore in both wild type and mutant embryos (Figure 1C). Although a mild effect could be missed in these qualitative analyses, these observations indicate that <u>CHE-14</u> is not necessary for the secretion and apical localization of <u>GRL-2</u> and <u>WRT-10</u>.

Another *ptr* gene, <u>*daf-6*</u>, has been reported to function redundantly with <u>*che-14*</u> in single-cell tubulogenesis, and loss of function of both genes causes synthetic larval lethality (Perens and Shaham 2005). However, embryos mutant for both genes were still able to secrete <u>WRT-10</u> (Figure 1C). Given the large number of *ptr* genes in <u>*C. elegans*</u>, it is possible that other *ptr* genes expressed in these epithelia act redundantly with <u>*che-14*</u> to transport <u>GRL-2</u> and <u>WRT-10</u>, or that a different *ptr* gene is specifically required. It is also possible that <u>CHE-14</u> transports different Hh-r proteins than those studied here, or perhaps <u>*C. elegans*</u>. Hh-r proteins may not rely on PTR proteins for secretion.

A recent report demonstrated that endocytosis of the Hh-r <u>GRL-7</u> specifically requires <u>PTR-18</u>, suggesting that the PTRproteins might regulate endocytosis or other aspects of Hh-r protein trafficking (Chiyoda et al., 2021). Emerging data also suggest that some Hh-r proteins work with specific PTR proteins in possible signaling roles; however, the pathways that connect them with downstream effects remain elusive (Riveiro et al., 2017; Lin and Wang 2017; Kume et al., 2019; Templeman et al., 2020; Chiyoda et al., 2021; Shi and Murphy 2023; Wang et al., 2023; Emans et al., 2023). Further work is needed to reveal the connection between these protein families in nematodes.

## Methods

## <u>*C. elegans*</u> culture and maintenance:

All strains were maintained at 20° C on nematode growth medium (NGM) seeded with <u>OP50</u> *E. coli* under standard conditions (Brenner 1974). Information from Wormbase was referenced for all experimental design (Sternberg et al., 2024). Strains used are listed in the Reagent table.

Confocal imaging of fluorescent proteins in elongating *C. elegans* embryos:

Images were captured using a Leica TCS DMi8 confocal microscope and Leica Application Suite X software (version 3.5.7.23225). This microscope was equipped with an HC PL APO 63X objective lens (Numerical Aperture 1.3). Entire embryos were imaged by collecting a series of 60-75, 0.33 µm Z-slices. SfGFP fusions were visualized using a 488 nm laser set to 3% power and emitted wavelengths between 493 and 578 nm were collected at a scanning speed of 700 hz with a HyD sensor. *C. elegans* embryos were picked at the 1.5fold stage of development and incubated at 20° C for the indicated number of hours prior to imaging. Samples were mounted with 10 mm levamisole in M9 buffer on pads of 2% agar noble and 2.5% sodium azide.

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## Reagents

Strain	Genotype	Source
<u>ML514</u>	<u>che-14(ok193</u> ) I	CGC, Michaux et al., 2000
<u>CB1377</u>	<u>daf-6(e1377</u> ) X	CGC, Perens and Shaham 2005
PHX7022	<u>wrt-10</u> (syb7022 [ <u>WRT-10</u> ::sfGFP]) II	SunyBiotech, Serra et al., 2024
PHX7064	<u>grl-2(</u> syb7064 [ssSfGFP:: <u>GRL-2]</u> ) V	SunyBiotech, Serra et al., 2024
UP4228	<u>che-14(ok193</u> ) I; <u>wrt-10</u> (syb7022) II	This work
UP4229	<u>che-14(ok193</u> ) I; <u>grl-2</u> (syb7064) V	This work
UP4274	<u>che-14(ok193</u> ) I; <u>wrt-10</u> (syb7022) II; <u>daf-6(e1377</u> ) X; csEx946( <u>che-14</u> +; unc- 119p::GFP)	This work. The <u>che-14</u> rescue transgene csEx946 was generated by co-injecting fosmid WRM0620dD06 [20 ng/µl] and pIM175 (unc-119p::GFP) [100 ng/µl].

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