

Loss of *dpy-2* and *dpy-9* has stage-specific effects on DBL-1 pathway signaling

Mohammed Farhan Lakdawala¹ and Tina L. Gumienny^{1§}

¹Department of Biology, Texas Woman's University, Denton, TX, 76204-5799

[§]To whom correspondence should be addressed: tgumienny@twu.edu

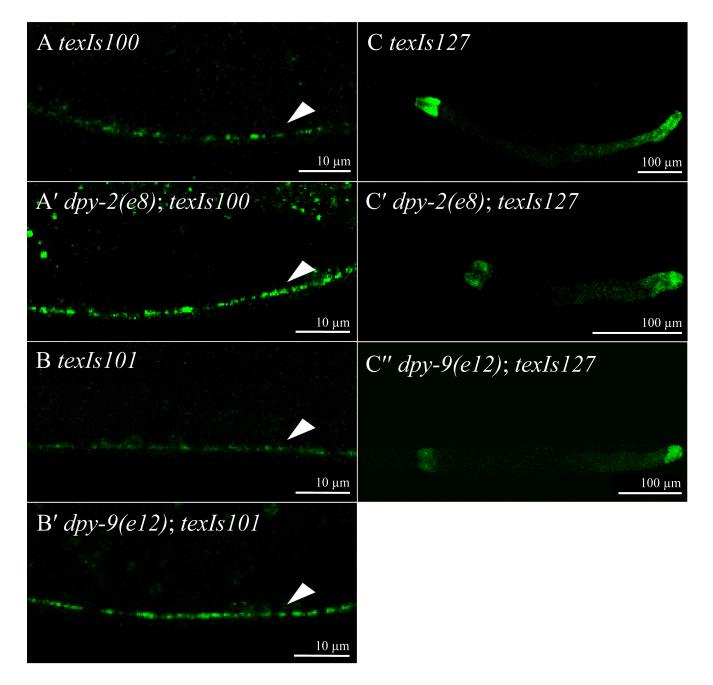


Figure 1: *dpy-2* or *dpy-9* loss-of-function mutations affect GFP::DBL-1 and DBL-1 pathway reporter fluorescence in L4 animals. Arrows point to GFP::DBL-1 fluorescent punctae in A and B. Representative images show that loss of *dpy-2* or *dpy-9* gene function is associated with increased GFP::DBL-1 fluorescence from *texIs100* or *texIs101* as shown in (A') and (B'), respectively. *dpy-2(e8)* and *dpy-9(e12)* mutants also have reduced *spp-9p::gfp* reporter activity compared to control (C), as shown in (C') and (C''), respectively.

Loading [Contrib]/a11y/accessibility-menu.js

12/9/2019 - Open Access

Description

Loss of some cuticle collagens negatively affects DBL-1 pathway signaling in a stage-dependent manner (Lakdawala et al. 2019; Madaan et al. 2019). We previously observed that in one-day old adult animals, loss of dpy-2 or dpy-9 had no effect on GFP::DBL-1 expressed from the *dbl-1* promoter (Beifuss and Gumienny 2012; Lakdawala et al. 2019). We also observed that expression of *spp-9p::qfp*, a reporter that is negatively regulated by the DBL-1 pathway, was not affected in one-day old adult animals (Roberts et al. 2010; Lakdawala et al. 2019). Post-embryonic expression of dpy-2 and dpy-9 is highest in L2 and L3, but low in L4 and even lower in young adults (Gerstein et al. 2010). Because cuticle secreted in one stage creates the cuticle in the next stage, this is consistent with the observation that loss of *dpy-2* and *dpy-9* has no effect on DBL-1 signaling in the adult (Hall and Altun 2008; Lakdawala et al. 2019). However, the DPY-2 and DPY-9 expression patterns led us to ask if DBL-1 signaling is affected at L4 by loss of dpy-2 or dpy-9. To our surprise, we found that dpy-2(e8) or dpy-9(e12) resulted in significant increases of GFP::DBL-1 fluorescence within DBL-1-secreting cells in L4 animals compared to control populations (Figure 1, Table 1). We also tested DBL-1 pathway reporter activity in these *dpy-2* and *dpy-9* mutants. Consistent with the increased GFP::DBL-1 fluorescence at L4, we observed significantly decreased fluorescence from the spp-9p::gfp reporter at L4 (Figure 1, Table 1). These results are consistent with DPY-2 and DPY-9 affecting DBL-1 signaling at the L4 stage but not at the adult stage. This suggests that these two collagens have a stage-specific effect on DBL-1 signaling, but this effect is normally inhibitory, as loss of *dpy-2* or *dpy-9* increased GFP::DBL-1 fluorescence and decreased *spp-9p*::GFP fluorescence.

Table 1: Effects of <i>dpy-2</i> and <i>dpy-9</i> gene mutations on GFP::DBL-1 and DBL-1 pathway reporter <i>spp-9p</i> ::GFP fluorescence						
Gene	Genotype	GFP::DBL-1 fluorescence % control ± 95% CI	P value	Genotype	<i>spp-9p</i> ::GFP % control ± 95% CI	P value
Animals at L4 stage						
control	texIs100	100±29.58	_	texIs127	100±7.94	_
control	texIs101	100±54.28	_	-	-	_
dpy-2	dpy-2; texIs100	155.47±55.58	0.0263	dpy-2; texIs127	80.26±10.62	0.0009
dpy-9	dpy-9; texIs101	212.94±98.06	0.0009	dpy-9; texIs127	84.37±9.56	0.0028
Animals at adult stage (data from (Lakdawala <i>et al.</i> 2019))						
control	texIs100	100±15.57	_	texIs127	100±11.47	_
control	texIs101	100±25.95	_	_	_	_
dpy-2	dpy-2; texIs100	115±52.15	0.5080	dpy-2; texIs127	107.04±12.20	0.2344
dpy-9	dpy-9; texIs101	95.02±29.01	0.7248	dpy-9; texIs127	100.29±10.24	0.9533

Methods

Request a detailed protocol

Nematode maintenance and imaging All the strains were maintained at 20°C on EZ media (Madhu et al. 2019). L4 animals were anesthetized using 1 mM levamisole hydrochloride (Sigma, St. Louis, MO) and imaged on a Nikon A1 confocal system (Nikon Instruments, Melville, NY). GFP::DBL-1 fluorescence was captured using a 60X objective and *spp-*9p::gfp fluorescence was captured using a 10X objective. The imaging conditions were optimized and kept constant between control and experimental samples. Nikon NIS Elements AR-5.02 software was used to quantify fluorescence intensities. Statistical analyses were performed using the unpaired t-test to compare control and experimental sample means. "% control \pm 95% CI" is the ratio of the indicated strain mean to the control strain mean \pm 95% confidence interval. n=10 for each strain imaged for the GFP::DBL-1 experiment, and n=15 for each strain imaged for the *spp-9p*::GFP experiment.

Reagents

Strains

Strains used in this study are:

TLG182 texIs100 [dbl-1::dbl-1:gfp; ttx-3p::rfp] IV

TLG205 texIs101 [dbl-1::dbl-1:gfp; ttx-3p::rfp] V

Loading [Contrib]/a11y/accessibility-menu.js



12/9/2019 - Open Access TLG697 texIs127 [spp-9p::qfp] X

TLG701 dpy-2(e8); texIs100

TLG702 dpy-9(e12); texIs101

TLG725 dpy-2(e8); texIs127

TLG724 dpy-9(e12); texIs127

Strains are available upon request.

Acknowledgments: We thank Cathy Savage-Dunn for helpful comments. We thank all our lab members for useful discussions. Some strains were obtained from the Caenorhabditis Genetics Center (CGC), which is funded by NIH Office of Research Infrastructure Programs (P40 OD010440). We thank WormBase.

References

Beifuss, K.K., and Gumienny, T.L. (2012). RNAi screening to identify postembryonic phenotypes in *C. elegans*. JoVE 60: e3442. PMID: 22353760.

Gerstein, M.B., Lu, Z.J., Van Nostrand, E.L., Cheng, C., Arshinoff, B.I. *et al.* (2010). Integrative analysis of the *Caenorhabditis elegans* genome by the modENCODE project. Science 330: 1775-1787. PMID: 21177976.

Hall, D., and Altun, Z. (2008). C. elegans Atlas. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York.

Lakdawala, M.L., Madhu, B., Faure, L., Vora, M., Padgett, R.W., and Gumienny, T.L. (2019). Genetic interactions between the DBL-1/BMP-like pathway and *dpy* body size-associated genes in *Caenorhabditis elegans*. Molecular Biology of the Cell, in press. https://www.molbiolcell.org/doi/pdf/10.1091/mbc.E19-09-0500 DOI: 10.1091/mbc.E19-09-0500

Madaan, U., Faure, L., Chowdhury, A., Ahmed, S., Ciccarelli, E.J., Gumienny, T.L., and Savage-Dunn, C. (2019). Feedback regulation of BMP signaling by *C. elegans* cuticle collagens. bioRxiv 686592.

Madhu, B., Salazar, A.E., and Gumienny, T.L. 2019 *Caenorhabditis elegans* egg-laying and brood-size changes upon exposure to *Serratia marcescens* and *Staphylococcus epidermidis* are independent of DBL-1 signaling. microPublication Biology. DOI: 10.17912/2r51-b476

Roberts, A.F., Gumienny, T.L., Gleason, R.J., Wang, H., and Padgett, R.W. (2010). Regulation of genes affecting body size and innate immunity by the DBL-1/BMP-like pathway in *Caenorhabditis elegans*. BMC Dev Biol 10: 61. PMID: 20529267.

Funding: This work was supported by NIH grants R01 GM097591, a TWU Chancellor's Research Fellowship to TLG, and internal funding by Texas Woman's University.

Author Contributions: Mohammed Farhan Lakdawala: Investigation, Formal analysis, Writing - original draft, Writing - review and editing. Tina L. Gumienny: Conceptualization, Resources, Funding acquisition, Project administration, Writing - review and editing.

Reviewed By: Anonymous

History: Received November 15, 2019 Accepted November 21, 2019 Published December 9, 2019

Copyright: © 2019 by the authors. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International (CC BY 4.0) License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Citation: Lakdawala, MF; Gumienny, TL (2019). Loss of *dpy-2* and *dpy-9* has stage-specific effects on DBL-1 pathway signaling. microPublication Biology. https://doi.org/10.17912/micropub.biology.000191