The *Arabidopsis* MYB5 transcription factor interacts with CASEIN KINASE2 BETA3 subunit in a yeast two-hybrid system

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Abstract

*Arabidopsis thaliana* MYB5 collaborates with TRANSPARENT TESTA GLABRA1 (TTG1) and basic-Helix-Loop-Helix (bHLH) transcription factors to regulate seed coat, trichome and root cell differentiation. Using a yeast two-hybrid system we show that the N-terminal region of MYB5 binds directly to the serine/threonine CASEIN KINASE2 BETA3 (CK2β3) subunit. Functions of the CASEIN KINASE2 (CK2) complex include facilitating phosphorylation of MYB transcription factors and cell cycle checkpoint regulatory proteins. Purified recombinant MYB5 protein was found to bind only weakly *in vitro* to the promoter of ALPHA/BETA ESTERASE/HYDROLASE4 (ABE4), a known MYB5 target gene. We propose that phosphorylation of MYB5 facilitated by the MYB5-CK2β3 interaction enhances MYB5 binding to its target gene promoters.

Figure 1. Yeast two-hybrid analysis of MYB5 interaction with CK2β3 protein kinase subunit and EMSA binding of MYB5 to the ABE4 promoter: (A) Schematic diagram showing three candidate MYB5 bait constructs containing the cDNA nucleotide sequences coding for MYB5 full length, N-terminal and C-terminal regions. The three MYB5 cDNA sequences were each fused in frame with GAL4BD in pGBKT7 plasmids. ADH1: ALCOHOL DEHYDROGENASE1, ATG: Methionine start codon, NLS: Nuclear localisation sequence, R2: MYB repeat 2, R3: MYB repeat 3. (B) Auto-activation screening of the candidate bait constructs. pMYB5FL::GAL4BD and pMYB5CT::GAL4BD colonies grew and turned blue while pMYB5NT::GAL4BD colonies failed to do so (no auto-activation). (C) Confirmation of protein-protein interaction between MYB5 and CK2β3 in yeast. Colonies transformed with pMYB5NT::GAL4BD and pCK2β3::GAL4AD grew while colonies transformed with pGAL4BD (empty pGBKT7 vector) and pCK2β3::GAL4AD failed to grow. Positive control: pVA3 (VA3 protein) and pTD1 (TD1 protein). (D) Electrophoretic mobility shift assay (EMSA) was used to determine MYB5 protein binding to a target promoter *in vitro*. The EMSA analysis showed weak binding affinity of AtMYB5trunc protein to an ABE4
(At2g23580) promoter sequence containing two MYB-related regulatory elements (MYBCORE and MYBLEPR binding motifs). Two weak band shifts were detected within the same probe indicating that MYB5 binds weakly to the enriched region containing the MYBCORE (CNGTTR) and MYBLEPR (GTAGTT) binding motifs (Li et al., 2020). Arrows indicate probe shift, - indicates labelled probe only, + indicates labelled probe plus AtMYB5 protein, T7 indicates labelled probe with the addition of T7 antibody. (E) Model for MYB5-CK2β3 interaction and MYB5 protein-DNA binding for activation of MYB5 target gene expression in plants. CK2 may facilitate phosphorylation of MYB5 directly or by recruiting additional kinase enzymes thereby assisting with MYB5-DNA binding to target gene promoters. bHLH: basic-Helix-Loop-Helix, (P): Phosphate group.

Description

The Arabidopsis (Arabidopsis thaliana) MYB5 (At3g13540) transcription factor (TF) forms transcriptional regulatory (MBW) complexes with basic-Helix-Loop-Helix (bHLH) and TRANSPARENT TESTA GLABRA1 (TTG1) (WD-repeat) TF proteins. These MBW complexes regulate seed coat, trichome and root cell differentiation via a multi-tiered transcriptional mechanism (Golz et al., 2018; Li et al., 2020). The Arabidopsis CK2β3 subunit is part of the CK2 holoenzyme which is a tetramer consisting of two regulatory β-subunits CK2β3 and CK2β4 subunits share 87% amino acid sequence identity and 92% sequence identity. CK2β3 and CK2β4 subunits share 87% amino acid sequence identity and 92% sequence similarity. Moreover, like CK2β3, CK2β4 has also been linked to the phosphorylation of CCA1 as over-expression of both genes leads to period-shortening of circadian clock-related gene expression (Perales et al., 2006). Although CK2β3 does not exhibit phosphorylation activity, it has been shown to facilitate CCA1 binding to the promoter of target gene CHLOROPHYLL A/B BINDING PROTEIN1 (CAB1) (Sugano et al., 1999). CK2β2 also interacts with the MYB-related LATE ELONGATED HYPOCOTYL (LHY) and YING YANG1 (YY1) via CK2β3 and phosphorylates LHY and YY1 proteins in vitro (Sugano et al., 1999; Wu and Li, 2017).

The transcriptional activities of Pinus taeda MYB4 (PtMYB4) and PtMYB46 are positively regulated by phosphorylation (Morse et al., 2009) although phosphorylation of TFs such as LTF1 (which regulates lignin biosynthesis in Populus) can result in its degradation (Gui et al., 2019). In contrast, DNA binding of Arabidopsis MYB15, a transcriptional repressor of cold signalling, is reduced by phosphorylation (Kim et al., 2017). While specific MYB5-CK2β3 interactions in planta are yet to be
determined, CK2B3 may facilitate direct MYB5 binding to its target gene promoters or may phosphorylate MYB5 indirectly by recruiting the CK2 catalytic α-subunits. The MYB5 N-terminal region contains several threonine and serine residues which are potential phosphorylation sites (Li et al., 1996) and phosphorylation of these residues may enhance MYB5 protein-DNA binding affinity. Alternatively, as MYB5-TTG1 and/or MYB5-bHLH interactions were not detected in the initial yeast two-hybrid screening, MYB5 may also require activation to facilitate TF protein-protein binding. TTG1 is subject to site-specific phosphorylation which regulates binding with TT2 (MYB123) to form some MBW complex combinations (Li et al., 2018). MYB5-C2B3 interactions and proposed phosphorylation may contribute to MYB5-bHLH-TTG1 (MBW) complex formation which, in turn, may further increase MYB5 protein-DNA binding affinity in vivo.

In the proposed model (Fig 1E), MYB5-bHLH-TTG1 complexes activate the expression of target genes by binding to their promoters. CK2B3 interacts with MYB5 transiently and recruits the CK2 holoenzyme to phosphorylate MYB5 (Fig 1E) resulting in more active MYB5-bHLH-TTG1 complexes thereby enhancing expression of MYB5 target genes.

Methods

Request a detailed protocol

Plasmid construction

For yeast two-hybrid (Y2H) analysis, MYB5 full length, C-terminal and N-terminal nucleotide sequences were PCR amplified from cDNA prepared from developing wild-type (Col) silique using Y2HFULLF/Y2FULLR [pMYB5FL], Y2HFULLF/Y2HNTR [pMYB5CT] and Y2HCTF/Y2FULLR [pMYB5NT] primer combinations, respectively. The MYB5 cDNA sequences were ligated into the pGBK7 yeast expression vector at the EcoR1 and BamHI restriction sites, generating pMYB5FL::GAL4BD, pMYB5CT::GAL4BD and pMYB5NT::GAL4BD vectors, respectively. For EMSA analysis, a truncated MYB5-encoding cDNA sequence (410 nucleotides) containing the MYB DNA-binding domain sequence with the truncation after the GIDPOTHK polyadenylation site was PCR amplified from wild-type (Col) seed cDNA using MYBPRTOF and MYBTRUNCR primers. Truncation of MYB proteins beyond the MYB domain does not appear to affect binding specificity (Li and Parish, 1995; Phan et al., 2011). The truncated MYB5 fragment was ligated into the pRSETA expression vector (Invitrogen) at XhoI and EcoR1 restriction sites and was fused to a 6X polyhistidine tag and a T7 leader peptide coding sequence and driven by the T7 promoter which was PCR amplified using T7F and PGADT7R primers.

Yeast Two-Hybrid analysis

The yeast two-hybrid (Y2H) assay was performed using the Matchmaker™ Gold Yeast Two-Hybrid system (Clontech) and all protocols were adapted from the Clontech Yeast Protocols Handbook and the Matchmaker™ Gold Yeast Two-Hybrid user manual. To determine the autoactivation of candidate bait constructs, Y2HGold cells were transformed with the three constructs and inoculated onto dropout (-trp) media containing x-α-gal and auribasidin A. Five technical replicates are presented (Fig 1B). pGAL4BD (empty pGBK7 vector) was used as a negative control and pVA3/pTD1 co-transformed cells were used as a positive control. The Mate & Plate™ Library – Universal Arabidopsis (Normalised) (Clontech) was used which consists of a cDNA library cloned into the GAL4AD vector (pGADT7, prey) and transformed into yeast strain Y187. Y2HGold yeast cells transformed with pMYB5NT::GAL4BD (bait) were mated with the Y187 cells expressing the Mate & Plate™ Library made from 11 Arabidopsis tissues. The mated cultures were plated onto the double dropout (~trp ~leu) media containing x-α-gal and auribasidin A. to identify bait and prey interaction. Yeast cells were then inoculated on double-dropout (DDO) medium (~leu/trp) plus x-α-gal and quadruple-dropout (QDO) (~ade/~his/~leu/~trp) medium plus x-α-gal and auribasidin A. Double-dropout medium (~leu/trp) selects for the presence of both pGADT7 and pGBK7. Figure 1C depicts one representative biological replicate of MYB5-CK2B3 binding. Four technical replicates are presented. In total, 3 out of 5 (60%) of the positive colonies from initial Y2H screening were confirmed as positive MYB5-CK2B3 interactions.

Electrophoretic Mobility Shift Assay (EMSA)

Recombinant MYB5 protein expressed in E. coli was purified using Ni-NTA Superflow columns (Qiagen) according to the manufacturer’s protocol. EMSA was performed using a non-radioactive protocol (adapted from Phan et al., 2011) and the protocol described by Li and Parish (1995). The ABE44COREF and ABE44CORER primers were designed to PCR amplify a probe of 150 to 200 nucleotides in length based on the promoter region enriched in ChIP analysis (Li et al., 2020). Protein-probe binding was performed using 400 ng total protein, 3 mL of binding buffer (20 mM NaCl, 5 mM MgCl2, 20 mM Tris [pH 8.0], 10% glycerol, 0.5 mM EDTA and 0.5 mM DTT) with 1 mL of poly d(I-C). The truncated MYB5 protein was expressed in E. coli and purified. SDS-PAGE and Coomassie Blue staining detected bands of approximately ~21kDa molecular weight in two independent replicates. Western Blot analysis using an anti-T7 epitope tag antibody detected ~21k kDa bands in each lane corresponding to the truncated MYB5 protein replicates. EMSA probes were labelled with digoxigenin via PCR using digoxigenin-labelled dUTP (Roche).
Reagents

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<tr>
<th>Primer name</th>
<th>Primer sequence (5’ to 3’ orientation)</th>
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References


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