Effect of the protease plasmin on *C. elegans* hyperactive DEG/ENaC channels MEC-4(d) and UNC-8(d)

Christina K. Johnson¹, David D. Miller² and Laura Bianchi¹§

¹University of Miami
²Vanderbilt University
§To whom correspondence should be addressed: lbianchi@med.miami.edu

Abstract

*C. elegans* MEC-4 and UNC-8 belong to the DEG/ENaC family of voltage-independent Na⁺ channels and have been implicated in mechanosensation and synaptic remodeling. MEC-4 and UNC-8 hyperactive mutants, designated (d) mutants, conduct enhanced currents and cause cell death due to uncontrolled influx of cations. We show here that MEC-4(d) but not UNC-8(d) currents are further potentiated by treatment with the protease plasmin and that this effect is dependent upon co-expression with the chaperon protein MEC-6. Mammalian DEG/ENaC channels are cleaved by plasmin in the channel finger domain and both MEC-4 and UNC-8 have a predicted plasmin cleavage site in this domain. We previously showed that MEC-4(d), but not UNC-8(d), currents are increased by co-expression with MEC-6, which interacts with the channel via the finger domain. We suggest that interaction of the channel subunit with MEC-6 may render the plasmin cleavage site more accessible. Given that *C. elegans* expresses a homolog of plasmin, these effects might be relevant *in vivo*.
Figure 1. Plasmin enhances MEC-4(d) but not UNC-8(d) currents in a MEC-6-dependent manner: Estimation plots by unpaired t-Test analysis of currents recorded in oocytes not treated and treated with plasmin (Calin-Jageman and Cumming 2019). Individual data points, means and 95% confidence intervals are shown. In each graph, on the right, the black circle and the error bars indicate the difference between the means and its 95% confidence interval. A: Currents recorded at -100 mV from oocytes expressing MEC-4(d)+MEC-2+MEC-6 not treated (light red) and treated (dark red) with 10 μg/ml plasmin, as indicated. N = 15, each. The difference between the means was 4.68 (71% increase), the lower and upper 95% confidence levels were 2.45 and 6.83 respectively (38% and 104% change, respectively). B: Same as panel A for oocytes expressing UNC-8(d)+MEC-2+MEC-6 not treated (light blue) and treated (dark blue) with plasmin. N = 7 and 5. The difference between the means was 0.94 (6% increase), the lower and upper 95% confidence levels were -1.50 and 3.38 respectively (-10% and 22% change, respectively). C: Same as in A for oocytes expressing MEC-4(d)+MEC-2 not treated (light red) and treated (dark red) with plasmin. N = 15 and 13 respectively. The difference between the means was 0.003 (0% change), the lower and upper 95% confidence levels were -0.391 and 0.398 respectively (-41% and 41% change; such large percentage changes are due to the relatively large variability of small currents). D: Same as panel A for oocytes expressing UNC-8(d)+MEC-2 not treated (light blue) and treated (dark blue) with plasmin. N = 6 and 5. The difference between the means was -0.87 (6% decrease), the lower and upper 95% confidence levels were -2.17 and 0.44 respectively (-27% and 15% change, respectively). E: Alignment of the finger domain of MEC-4 and UNC-8 containing the predicted plasmin sites (yellow background). F: Schematic representation of a DEG/ENaC channel subunit based on its crystal structure (Jasti, Furukawa et al. 2007). Alpha helixes are shown as cylinders and beta sheets are represented as flat arrows. The finger domain (purple) contains the plasmin recognition site (yellow box). The shaded area around transmembrane domains 1 and 2 (TM1 and TM2) represents plasma membrane.

Description


Potentiation of the mammalian α, γ, and δ subunits of the DEG/ENaC ENaC subfamily can be also achieved via proteolytic cleavage of the extracellular domain (Kleyman and Eaton 2020). This cleavage results in the release of inhibitory peptides and probably the conformational change of the channel that leads to larger whole-cell currents. DEG/ENaC proteolytic processing begins in the Golgi by the Golgi resident protease furin (Hughey, Mueller et al. 2003, Hughey, Bruns et al. 2004) and is completed once the channel arrives at the cell surface by transmembrane and extracellular proteases such as plasmin (Passero, Mueller et al. 2008). While the effect of proteases on mammalian DEG/ENaCs has been extensively studied, little is known about whether nematode DEG/ENaC subunits are sensitive to these enzymes.

We coexpressed MEC-4(d) with stomatin-like protein MEC-2 and paraoxonase-like protein MEC-6, and, in a separate experiment, UNC-8(d) with MEC-2 and MEC-6 in *Xenopus* oocytes and recorded currents using the two-electrode voltage-clamp technique. In the same batch of oocytes, we compared current amplitudes recorded at -100 mV in cells treated with 10 μg/ml of plasmin (in ND96) for 30 minutes with currents recorded in mock-treated cells (ND96). We found that MEC-4(d) currents were nearly doubled in plasmin-treated oocytes (Fig. 1A) (6.54 +/- 0.68 and 11.23 +/- 0.79 for mock and plasmin-treated cells respectively, mean +/- SE, n=7 and 5). Mammalian ENaCs are cleaved by plasmin in the channel finger domain where this protease has at least one recognition site (Passero, Mueller et al. 2008). We thus used the SitePrediction website (https://www.dmbr.ugent.be/prx/bioit2-public/SitePrediction/) to predict potential plasmin cleavage sites in the finger domains of MEC-4 and UNC-8 (Fig. 1E and F). We found that both proteins contain at least one potential plasmin cleavage site in the finger domain (Fig. 1E, highlighted in yellow). This suggests that lack of effect of plasmin on UNC-8(d) is not due to lack of potential recognition sites for plasmin in UNC-8.

MEC-6 is a paraoxonase-like protein (Chelur, Ernstrom et al. 2002) with chaperone function (Chen, Bharill et al. 2016) that increases the number of functional MEC-4 channels at the cell surface (Brown, Liao et al. 2008, Chen, Bharill et al. 2016). We
recently showed that MEC-6 mediates MEC-4(d) regulation via interaction with the channel finger domain (Matthewman, Johnson et al. 2018). We also showed that MEC-6 does not enhance UNC-8(d) currents in oocytes. We thus postulated that the effect of plasmin on MEC-4(d) might depend on MEC-6 and that the lack of effect on UNC-8 might be related to the fact that MEC-6 does not change UNC-8 currents in Xenopus oocytes. To test this hypothesis, we repeated the experiments in oocytes injected with only MEC-4(d) and MEC-2 or with only UNC-8(d) and MEC-2, but without MEC-6. As previously reported, MEC-4(d) currents are much smaller in the absence of MEC-6 (Chelur, Ernstrom et al. 2002). More importantly, we found that plasmin does not increase MEC-4(d) currents in oocytes that do not express MEC-6 (Fig. 1C) (0.95 +/- 0.14 and 0.95 +/- 0.13 for mock and plasmin treated cells respectively, mean +/- SE, n=13 and 15). UNC-8(d) currents remain unaffected by plasmin treatment (Fig. 1D) (14.32 +/- 1.09 and 13.44 +/- 0.65 for mock and plasmin treated cells respectively, mean +/- SE, n=6 and 5). Taken together, these results support the idea that chaperon MEC-6 is needed for enhancement of MEC-4(d) current amplitude by protease plasmin. Interestingly, Brown and colleagues showed that activation of MEC-4(d) by extracellular chymotrypsin is independent of both MEC-2 and MEC-6 (Brown, Liao et al. 2008). Collectively, these data suggest that *C. elegans* DEG/ENaC channels might be sensitive to different extracellular proteases based, not only on channel primary sequence but also on its quaternary structure. These results also suggest that chaperon MEC-6 may help expose potential cleavage sites. Given that *C. elegans* expresses *svh-1*, the homolog of mammalian plasmin precursor plasminogen, we suggest that these effects might be relevant *in vivo* both in physiological conditions and when DEG/ENaC channels are hyperactivated (Hisamoto, Li et al. 2014).

**Methods**

**Request a detailed protocol**

**Molecular biology**

Plasmids encoding MEC-4(d) and MEC-2 were a gift from the Chalfie lab (Goodman, Ernstrom et al. 2002). The MEC-6 and UNC-8(d) cDNA were cloned in a PGEM vector and were published previously (Bianchi, Gerstbrein et al. 2004, Wang, Matthewman et al. 2013, Matthewman, Miller-Fleming et al. 2016, Matthewman, Johnson et al. 2018). cRNAs were synthesized using T7 mMESSAGE mMACHINE kit (Invitrogen) following manufacturer’s protocols. cRNAs were purified, run on denaturing agarose gels for size and integrity verification, and quantified spectrophotically.

**Xenopus oocytes expression and electrophysiology**

Stage V-VI defolliculated oocytes from *Xenopus laevis* were purchased from Ecocyte Bioscience US LLC (Austin, Texas) and injected with 10 ng/oocyte of each cRNAs except MEC-6 cRNA, which was injected at 2 ng/oocyte. Injected oocytes were incubated in ND96 medium consisting of 96 mM NaCl, 2 mM KCl, 1.8 mM CaCl_2, 1 mM MgCl_2, and 5 mM HEPES, pH 7.5, supplemented with penicillin and streptomycin (0.1 mg/ml) and 2 mM Na-pyruvate at 20°C. The next day the medium was changed to ND96 plus 500 µM amiloride to prevent cell death (Goodman, Ernstrom et al. 2002). Whole-cell currents were measured three days after the injection using a two-electrode voltage-clamp amplifier (GeneClamp 500B; Axon Instruments) at room temperature. Electrodes with resistance between 0.3–0.7 MΩ were filled with 3 M KCl. To measure MEC-4(d) currents, oocytes were perfused with a physiological NaCl solution containing (mM): 100 NaCl, 2 KCl, 1 CaCl_2, 2 MgCl_2, and 10 HEPES, pH 7.2 (NaOH). Due to the strong block by divalent cations (Wang, Matthewman et al. 2013), to measure UNC-8(d) currents a divalent cation–free NaCl solution containing (mM): 110 NaCl, 2 KCl, 1 EGTA, and 10 HEPES, pH 7.2 (NaOH) was used. To elicit the whole-cell currents, the oocyte membrane was clamped at −30 mV and stepped from −160 to −100 mV in 20 mV increments. Current amplitude was measured at -100 mV and 300 ms. We used the pCLAMP suite of programs (Axon Instruments) for data acquisition and analysis. Currents were filtered at 200 Hz and sampled at 1 kHz. Prism was used to plot data and for statistical analysis.

**Reagents**

Plasmin was purchased from Calbiochem (cat # 527624-10U).

**Acknowledgments:** We thank the members of the Bianchi laboratory for technical support.

**References**


Wang, Y., C. Matthewman, L. Han, T. Miller, D. M. Miller, 3rd and L. Bianchi (2013). "Neurotoxic unc-8 mutants encode constitutively active DEG/ENaC channels that are blocked by divalent cations." J Gen Physiol 142(2): 157-169. DOI: 10.1085/jgp.201310974

Funding: This work was supported by NIH Grants R01s NS070969 and NS105616A1 awarded to Laura Bianchi and NS106951 awarded to David M. Miller.

Author Contributions: Christina K. Johnson: Investigation, Writing - review and editing. David D. Miller: Conceptualization, Writing - review and editing. Laura Bianchi: Conceptualization, Investigation, Writing - original draft, Writing - review and editing.

Reviewed By: Anonymous

History: Received May 20, 2021 Revision received June 17, 2021 Accepted June 21, 2021 Published June 21, 2021

Copyright: © 2021 by the authors. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International (CC BY 4.0) License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Citation: Johnson, CK; Miller, DD; Bianchi, L (2021). Effect of the protease plasmin on C. elegans hyperactive DEG/ENaC channels MEC-4(d) and UNC-8(d). microPublication Biology. https://doi.org/10.17912/micropub.biology.000412