Endogenous localization of TOP-2 in *C. elegans* using a C-terminal GFP-tag

Christine K. Rourke¹, Darline Murat¹, Tyler J. Hansen² and Aimee Jaramillo-Lambert¹§

¹Department of Biological Sciences, University of Delaware, Newark, Delaware 19716
²Currently-Department of Biochemistry, Vanderbilt University School of Medicine, Nashville, Tennessee 37205
§To whom correspondence should be addressed: anjl@udel.edu

Abstract

To investigate the dynamic localization of Topoisomerase II in live *C. elegans* we have generated a C-terminally GFP-tagged version of TOP-2 at the endogenous locus. We found that TOP-2::GFP localizes in a similar pattern to the previously published TOP-2::3XFLAG strain and does not disrupt the meiotic chromosome segregation functions of this enzyme.

![Image of localization](image-url)

**Figure 1. Endogenous TOP-2 localization using a GFP-tag:** (A-F) Live imaging of the *top-2::gfp* *C. elegans* strain using confocal microscopy. (A) A complete representation of TOP-2 localization in the germ line of *top-2::gfp* worms. Scale bar= 50 µm. (B, C) TOP-2 localization in the head and tail, respectively, of *top-2::gfp* worms. Scale bar= 50 µm (D) TOP-2 localization during pachytene of *top-2::gfp* worms. White arrowheads point to examples of TOP-2::GFP foci. Scale bar= 5 µm. (E) TOP-2 localization in -1 and -2 oocytes in *top-2::gfp* worms. Scale bar= 5 µm. (F) TOP-2 localization in developing embryos in *top-2::gfp* worms. Scale bar= 50 µm. (G) Brood size comparison of endogenously tagged *top-2::gfp* to wild type (N2). (H) Percent embryonic viability from the *top-2::gfp* strain compared to wild type (N2).

Statistics were conducted using a two-tailed Student’s T-test, n.s. indicates not significant, *p=0.0001.

**Description**

Topoisomerase II is a homodimeric, ATP-dependent enzyme that is conserved across species (Dong and Berger 2007; Nitiss 2009; Kim, Lee, and Koo 2000; Pommier et al. 2016; Vos et al. 2011). Topoisomerase II (Topo II) is expressed in all eukaryotic cells and is essential for many cellular processes including replication, transcription, chromosome structure, and chromosome segregation during mitosis (Nitiss 2009). In meiosis, Topo II is required for proper homologous chromosome segregation at the first meiotic division (Jaramillo-Lambert et al. 2016;
To determine whether the addition of the GFP protein tag impacts the function of TOP-2 we examined the effects on embryonic viability and brood size in the top-2::gfp worm strain. Overall brood size of the top-2::gfp worm strain was slightly reduced in top-2::gfp worms [243 (wild type, N2) vs. 194 (top-2::gfp), p=0.0001, Figure 1G]. The slight reduction in brood size in top-2::gfp is consistent with the top-2::3Xflag (Avg brood=141.5) strain we generated in a previous study (Jaramillo-Lambert et al. 2016). The reduction in brood size could either be due to minor disruptions in TOP-2 function in mitotically dividing cells within the proliferative zone of the germ line or due to TOP-2 defects in sperm production. As we aim to use the TOP-2::gfp worm strain to study the role of TOP-2 in male meiosis and spermatogenesis, future experiments will help resolve the two possibilities. However, embryonic viability of top-2::gfp worms is not significantly different from wild-type embryonic viability [99.1% (wild type) vs. 98.9% (top-2::gfp), p=0.79, Figure 1H]. In addition, we monitored the TOP-2::gfp strain for a high incidence of males phenotype, which is the consequence of X-chromosome nondisjunction. X-chromosome nondisjunction in wild-type worms is rare with less than 0.3% male progeny in self-fertilizing hermaphrodites (Hodgkin, Horvitz, and Brenner 1979; Lui and Colaiácovo 2013). The percent of males produced in top-2::gfp was not significantly different than wild-type worms (0% [wild type] vs. 0.6% [top-2::gfp], p=0.06). These results indicate that the addition of the GFP protein tag to TOP-2 is not disrupting the overall meiotic functions of this important enzyme. We conclude that the endogenous C-terminally tagged TOP-2::GFP creates a functional TOP-2 protein that is suitable for endogenous protein localization studies.

**Methods**

**Request a detailed protocol**

Generation of C-terminally tagged TOP-2::GFP via CRISPR-Cas9: A C-terminally meGFP-tagged version of TOP-2 (top-2::gfp) was created via the direct delivery method (Paix et al. 2017) using dpy-10 as a co-CRISPR marker (Arribere et al. 2014). The injection mix contained Cas9 protein (10 mg), dpy-10 CRISPR RNA (crRNA) (3.2 mg), dpy-10(c640) repair oligonucleotide (0.2 mg), universal trans-activating crRNA (tracrRNA) (20 mg, Dharmacon, GE Life Sciences), and a crRNA targeting the c-terminal sequence of top-2 (CACCGCGUCGAGACUCUGCAGUGUUUAAGCUUAGCUUUG, 8 mg). The repair template was generated by PCR using primers containing meGFP (5’-TTACGACGTGATCAGATCGATCCAGCCAGGAGGAGCTTCTCCCGC–3’) and top-2 reverse sequences (5’-GGGAAGAAAGAAAGAAATAATCATACCTAAAGGAAGGAGCTTGGAGAAAACAAATTGATTTTGATCCTGCTCCCTCAGGC–3’). The bold regions in the primers refer to the meGFP sequence. Edits were confirmed through PCR using a forward primer (5’-CGACAGCTGATGAGAGCTTCTCCCGC–3’) and a reverse primer (5’-CTCCTCCCTGAGTGATGAGCTTCTCCCGC–3’). The crRNA was designed to have a low possibility of off-target sites using http://crispr.mit.edu. Two independent strains were created (AG330 and AG332). Strain AG332 was the focus of this study.

Live imaging was conducted by placing 10 worms on a slide with a fresh 2% agarose pad and 10 µl of anesthetic (2 mM tetrathiomolybdate in M9 buffer). A glass coverslip was then placed on top of the worms and agarose pad and the space between the agarose pad and the edges of the coverslip was filled by pipetting M9 under the coverslip. Live images were taken on a Zeiss LSM880 confocal microscope using either the 40x (Figure 1A, B, C, & F) or the 63x (Figure 1D & E) objectives. Image processing and analysis was done on Fiji Is Just ImageJ (Schindelin et al. 2012).

Embryonic viability assays for the wild type (N2) and top-2(av87) (top-2::gfp) strains were conducted at 20°C. Both strains were grown on MYOB petri dishes inoculated with the E. coli (OP50). Ten L4 hermaphrodite larvae of each strain were put onto separate plates and allowed to grow and lay eggs for 16-24 h (one worm per plate). After each 24 h period, the adult (parent) worm was transferred onto a fresh plate until no additional embryos were produced. The total number of progeny (larvae and dead embryos) were counted one day after the parent was transferred. Three independent trials were conducted for each strain.

Brood sizes were determined by placing a single L4 hermaphrodite for both strains [N2 and top-2(av87) (top-2::gfp)] on a 35 mm MYOB plate spotted with E. coli (OP50). Each hermaphrodite was allowed to lay embryos for 24 h at 20°C. After 24 h the hermaphrodite was transferred to a fresh plate until no more embryos were produced (two additional transfers). Total brood size was calculated by adding together live larvae and dead embryos produced from a single hermaphrodite.

**Reagents**

AG332 top-2(av87) [top-2::gfp] is available from the Jaramillo-Lambert Lab. N2 is available from the CGC.

**Acknowledgments:** We would like to thank Andy Golden for his support in starting the study of C. elegans TOP-2 during meiosis.
References


Funding: This work was supported by a grant R03HD098244 from the National Institutes of Health to A.J.L and by the Institutional Development Award (IDeA) from the National Institute of Health’s National Institute of General Medical Sciences under grant number P20GM103446. Microscopy access was supported by grants from the NIH-NIGMS (P20 GM103446), the NSF (IIA-1301765) and the State of Delaware.

Author Contributions: Christine K. Rourke: Investigation, Methodology, Writing - original draft, Formal analysis. Darline Murat: Data curation, Formal analysis. Tyler J. Hansen: Methodology, Validation. Aimee Jaramillo-Lambert: Conceptualization, Funding acquisition, Supervision, Writing - review and editing.

Reviewed By: Anonymous

History: Received May 16, 2021 Revision received May 24, 2021 Accepted May 25, 2021 Published May 28, 2021

Copyright: © 2021 by the authors. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International (CC BY 4.0) License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.