Anti-oxidant MitoQ rescue of AWB chemosensory neuron impairment in a C. elegans model of X-linked Adrenoleukodystrophy

Sanjib Guha1§*, Aurora Pujol2,3,4* and Esther Dalfo5,6,7§*

1University of Rochester, Department of Anesthesiology & Perioperative Medicine, Rochester, NY
2Neurometabolic Diseases Laboratory, Bellvitge Biomedical Research Institute (IDIBELL), 08908 L’Hospitalet de Llobregat, Barcelona, Catalonia, Spain
3Center for Biomedical Research on Rare Diseases (CIBERER), ISCIII, Madrid, Spain.
4Catalan Institution of Research and Advanced Studies (ICREA), Barcelona, Catalonia, Spain.
5Faculty of Medicine, University of Vic-Central University of Catalonia (UVic-UCC), 08500 Vic, Spain
6Institut de Neurociències, Universitat Autònoma de Barcelona, 08193 Bellaterra, Spain
7Departament de Bioquímica i Biologia Molecular, Universitat Autònoma de Barcelona, 08193 Bellaterra, Spain
§To whom correspondence should be addressed: sanjib_guha@urmc.rochester.edu; esther.dalfo@umedicina.cat
*These authors contributed equally.

Abstract

X-linked Adrenoleukodystrophy (X-ALD) is a neurometabolic disorder caused by a defective peroxisomal ABCD1 transporter of very long-chain fatty acids (VLCFAs). We have characterized a nematode model of X-ALD with loss of the pmp-4 gene, the worm orthologue of ABCD1. These mutants recapitulated the key hallmarks of X-ALD and importantly mitochondria targeted antioxidant MitoQ prevented axonal degeneration and locomotor disability. In this study, we further demonstrated that the AWB chemosensory neuron of the pmp-4 mutant worm is defective, both in morphology and function. Interestingly, MitoQ could rescue both the phenotypes. Collectively, our results suggest that C. elegans’ chemosensation might provide a novel setting for exploring peroxisomal disease related disorders.
Figure 1. Antioxidant MitoQ rescue of AWB impairment in pmp-4 mutant worms: A. Representative DIC-Nomarski picture of an L4 worm head. B-F: DiI staining. The DiI stained neurons is delimited by the black square. B. Wild type animal in which all neurons stained by the DiI are visible and named. C. DiI staining of the pmp-4(ok396) mutant animal. The white dashed circle indicates limited staining of the AWB neuron. D. DiI staining of the rescue strain EDC010 pmp-4(ok396);ibbEx42[Ppmp-4::pmp-4::gfp]. The white dashed circle indicates the faint staining of the AWB neuron. E, F. Wild
X-linked adrenoleukodystrophy (X-ALD) is a rare neurometabolic disease characterized by inflammatory demyelination in the brain and axonal degeneration in the spinal cord. The disease is caused by mutations in the ATP binding cassette subfamily D member (ABCD1) gene, which encodes the peroxisomal transporter of very long chain fatty acids (VLCFAs) (Ferrer et al., 2015). Considering the possibility of establishing our model as a new platform for the investigation of ciliopathies and willing to go further along this new peroxisome-cilia association (Miyamoto et al., 2020), we deeply investigated the functionality of the amphid organ located in the head of the worm, composed of glia-like cells surrounding the chemosensory neurons (Stout et al., 2017). In addition, peroxisomes also supply cholesterol to primary cilia, non-motile antenna-like protrusions that picks up signals required for embryonic development and adult tissue maintenance (Miyamoto et al., 2020). This links peroxisomal disorders with another group of human diseases called ciliopathies, characterized by cilia dysfunction (Waters et al., 2011).

Human ciliopathies have been investigated in C. elegans by studying the functionality of the amphid organ, the chemosensory organ located in the head of the worm, composed of glia-like cells surrounding the chemosensory neurons (Stout et al., 2014; Shaham et al., 2015). Considering the possibility of establishing our model as a new platform for the investigation of ciliopathies and willing to go further along this new peroxisome-cilia association (Miyamoto et al., 2020), we deeply investigated the functionality of the amphid organ in the C. elegans model of X-ALD, constituted by the VC189 pmp-4(ok396) strain. In our recent study, we used two different approaches. First, the lipophilic dye 1,1’-Diiodotridecyl-3,3,3’,3’-Tetramethylindocarbocyanine Perchlorate (‘DiI’; DiIC18(3), named DiI from now on, was used to visualize the dye filling behavior associated with chemosensory neurons functionality in the amphid organ (Schulz et al., 2012). Second, we used reporter strain named (CX3877 kyls156 [Pstr-1::odr-10::GFP]) to visualize the structural morphology of the specific chemosensory neuron, Amphid Wing B (AWB) (Olivier Mason et al., 2013). kyls expresses GFP-labeled ODR-10 in STR-1 (a Seven Transmembrane Receptor involved in maintaining cell to cell connection) expressing cells (Mukhopadhyay et al., 2008; Troemel et al., 1997). Results obtained with the DiI stain, showed that six pairs of neurons in the head amphid sensory organs fill with DiI in wild-type animals (Fig. 1A, B). However, we found that inactivation of PMP-4 caused failure of dye uptake, specifically in the AWB neuron, but had no effect on the adjacent DiI sensitive neurons (Fig 1C). Interestingly, this dye-filling defect of pmp-4 mutants was no longer detected by the rescue strain, containing PMP-4::GFP driven by the pmp-4 promoter and injected in pmp-4(ok396) animals; pmp-4 (ok396) [Pppm-4::pmp-4(cDNA)::GFP] (and named rescue strain from now on for clarity) (Fig 1D) thus confirming the specificity of this dye-filling phenotype. Our previous study demonstrated that mitochondrial ROS were responsible of X-ALD associated phenotypes (Coppa et al., 2020). Accordingly, we hypothesized and further investigated the in vivo ability of the mitochondria-targeted antioxidant, MitoQ, to protect against ROS-induced chemosensory neuronal damage in the pmp-4 mutant animals. Worms were treated separately with the MitoQ (at 5 µg/ml) and the DiI uptake by the AWB neuron of the anti-oxidant treated worms were analyzed. Astonishingly, AWB DiI staining was rescued in all MitoQ treated pmp-4 mutant animals (Fig 1 E, F, and G).

Dil dye-filling defect are frequently associated with defects in cilia or dendrite morphology (Olivier Mason et al., 2013; Ou et al., 2007) and mutants with defects in cilia or dendrite morphology frequently exhibit dye-filling defects in some or all dye-
filling neurons (Olivier Mason et al., 2013). To determine whether the observed dye-filing phenotype in the \textit{pmp-4} mutant animals reflected defects in the neuronal morphology, we studied the AWB cilia structure by crossing the reporter strain \textit{kyIs156 [Pstr-1:: odr-10::GFP],with pmp-4(ok396)} mutant animals (Fig 1H). Generally all AWB cilia in wild type animals possess the characteristic Y-shaped structure containing two branches of different lengths (Fig 1H), the distal ends of which exhibit an irregular morphology and exhibit some animal to animal variability in the lengths of each cilial branch, depending upon which temperature they are grown and in which conditions (presence or absence of food) (Mukhopadhyay et al., 2008).

However, \textit{pmp-4} mutant animals showed both cilia branches slightly shorter than in wild-type animals (Fig. 1K, L) thus suggesting an underlying correlation between failures of DiI uptake with the regulation of cilia length (Fig. 1I, 1K, L). Interestingly, like the dye-filing defect, the AWB cilia branch length defects were recovered in the \textit{pmp-4} rescue strain (Fig. 1K and L). Therefore PMP-4 is essential to maintain AWB cilia morphology. As we demonstrated in our previous manuscript how MitoQ rescued X-ALD associated phenotypes (Coppa et al., 2020), here also we observed that the same concentration of MitoQ also rescued the AWB distal cilia morphology in \textit{pmp-4} mutant worms (Fig 1 J, K, L). Finally, the functional behavior associated with the AWB neuron was measured by odorant assay with nonanone, which generally \textit{C. elegans} avoid (Troemel et al., 1997; Tanimoto et al., 2017). Odorant index in \textit{pmp-4} mutant worms was decreased in comparison to wild types, and MitoQ treated animals showed a recovery in this index, as well (Fig 1N). Thus, through mitochondrial antioxidant treatment, we have demonstrated that MitoQ suppresses ROS-induced chemosensation defects in the \textit{pmp-4 (ok396)} worms.

Apart from the X-ALD specific phenotypes described in Coppa et al., 2020, in this study we have discovered that \textit{pmp-4} mutant worms are also defective in AWB chemosensory neuron cilia structural integrity which can lead inadequate function and defective staining. This phenotype can be regulated by mitochondrial ROS, since the mitochondria specific anti-oxidant MitoQ completely rescues all the defective phenotypes. Collectively, our results suggest that \textit{C. elegans} chemosensation phenotype can be used as a platform for the investigation of human ciliopathies in which peroxisomes are involved, and provides another setting to explore peroxisomal-related disorders.

**Methods**

*Request a detailed protocol*

\textit{C. elegans} strains growth and maintenance

Nematodes were maintained at 20\(^\circ\)C on Nematode Growth Media (NGM) plates made with Bacto Agar (BD Biosciences). The plates were seeded with live \textit{E. coli} OP50-1 bacterial strain (cultured overnight at 37\(^\circ\)C at 220 rpm) and allowed to grow overnight, as previously described in (Brenner S, 1974). For experimental assays, after synchronization by standard procedure with sodium hypochlorite, 4th larval stage (L4) hermaphrodites (characterized by the appearance of a “Christmas tree vulva”) were selected and used for all the experiments.

**Dye Filling Assay**

DiI: 1,1’- Dioctadecyl-3,3,3’,3’- Tetramethylindolo carbocyanine Perchlorate (Aldrich) staining was performed as previously described in (Schultz et al., 2012 and Tong et al., 2010). Briefly, a stock dye solution of 2 mg/ml DiI in dimethyl formamide was stored at -20\(^\circ\)C in a tube wrapped in foil to avoid oxidation. L4-staged well-fed worms from a plate were transferred into an eppendorf tube with 1 ml M9, into which 5 ul Dil solution from the stock was added. Tubes were shielded from light with aluminum foil and incubations were carried out at room temperature on a slow shaker for overnight. Next day, worms were mounted on a thick layer of half-dried agar (3%) pad on microscopic glass slides and subjected to confocal microscopy.

**Confocal Microscopy**

For ciliary morphology, animals were grown at the appropriate temperature were mounted on agarose pads set on microscopic slides and anaesthetized using 50 mM sodium azide in water (Sigma). Confocal images were acquired using a Leica spectral confocal microscope equipped with 63X objectives. Cilia length and morphology measurements were performed using ImageJ software (National Institutes of Health), after Z-stacking all the images, as described in Guha et al., 2020(b).

**MitoQ Assay**

For rescue experiments worms were fed with MitoQ (mitoquine mesylate): 10-(4,5-dimethoxy-2-methyl-3,6-dioxycyclohexa-1,4-dien-1-yl)decyltriphenylphosphonium methanesulfonate, as described in (Ng et al., 2014; Coppa et al., 2020). One milligram of MitoQ\(_{10}\) was dissolved in 1 ml of distilled water and 250 ul (1mg/ml) was added to autoclaved and cooled to 60\(^\circ\)C NG agar medium (total 50 ml). Synchronized L1 worms were placed on the MitoQ and normal NGM agar plates and grown for 48 hours till they reach the L4 stage. In parallel the same experiment was performed in plates without MitoQ as a control. Finally, worms were washed of the plates with M9 buffer and similar DiI or confocal experiment was performed as mentioned before.
Odortaxis assays

Avoidance assays for 2-nonanone was performed on square plates as described previously (Troemel et al., 1997). Plates were divided into six sectors labeled A-F. One microliter each of odorant (10% Nonanone) and 1 M NaN3 were added in two spots in sector A, and 1 μl each of control diluent (absolute ethanol) and 1 M NaN3 were added in two spots in sector F. An avoidance index (AI) or odortaxis index was calculated as [(number of animals in sectors A and B) − (number of animals in sectors E and F)]/ Total number of animals in all six sectors of the plate.

Reagents

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Name</th>
<th>Source</th>
<th>Comment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wild type (WT)</td>
<td>N2</td>
<td>CGC</td>
<td></td>
</tr>
<tr>
<td>pmp-4(ok396) IV</td>
<td>VC189</td>
<td>CGC</td>
<td></td>
</tr>
<tr>
<td>pmp-4(ok396) IV</td>
<td>EDC1</td>
<td>This study</td>
<td>10X outcrossed</td>
</tr>
<tr>
<td>kyls156 [Pstr-1::odr-10::GFP]</td>
<td>CX3877</td>
<td>CGC</td>
<td>GFP expression, specifically in the AWB neuron</td>
</tr>
<tr>
<td>kyls156 [Pstr-1::odr-10::GFP]; pmp-4</td>
<td>EDC61</td>
<td>This study</td>
<td></td>
</tr>
<tr>
<td>ibbEx42[Ppmp-4::pmp-4::GFP::(pmp-4)3'UTR; rol-6(su1006)]</td>
<td>EDC8</td>
<td>This study</td>
<td></td>
</tr>
<tr>
<td>pmp-4(ok396) IV; ibbEx42[Ppmp-4::pmp-4::GFP::(pmp-4)3'UTR; rol-6(su1006)]</td>
<td>EDC10</td>
<td>This study</td>
<td>rescue strain used to confirm the specific role of PMP-4 in ciliary abnormalities</td>
</tr>
</tbody>
</table>

All the strains can be ordered on request from Prof. Dalfo’s lab which is situated in the Institut de Neurociències, Autonomous University of Barcelona, Bellaterra Campus, Catalonia, Spain.

GFP fluorescence was used to guide selection of progenies, and PCR genotyping was used to confirm homozygosity with primers specific to the ok396 deletion, including: 5´-TCGGTAATCCCTTGTCTTCTC-3´ (forward), 5´-CGGAGGTCATCAGGTTTGTT-3´ (reverse) and 5´-ACTCCGAAGCCGATGAAATT-3´ (deletion).

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References


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