Variation between nematodes in a multi-sensory behavioral assay

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Description

An environment is often represented by numerous sensory cues. In order to better survive, an animal often needs to detect and process sensory cues simultaneously to make an appropriate behavioral decision. The Caenorhabditis elegans (C. elegans) genome encodes homologues of a significant number of molecules expressed in mammalian brains, which allows characterizing of the molecular and circuit basis for multi-sensory behavior during decision-making (Bargmann, 1998). In addition, studies have demonstrated various genes and neurons in behavioral differences previously observed in sensory behavior and decision-making when comparing different organisms, such as, nematodes. These variations in behavior involve differences in neuronal signals and neurons. For example, catecholamine signaling, neuropeptide Y-like receptors and pheromone chemoreceptors (Debono and Bargmann, 1998, Srinivasan et al., 2008; Bendesky et al., 2010, Mcgraph et al., 2011). In this study, we investigate the behavioral differences across various species of nematodes, by examining
multiple types of *Caenorhabditis* species and how they may differ in a multi-sensory behavioral assay (Harris et al., 2019). The multisensory behavioral assay involves examining how different species of nematodes that are exposed to conflicting cues, behave when compared to the standard wild type, *C. elegans* Bristol N2 worms. We use a multi-sensory behavior paradigm to address these questions to determine any difference in ‘2-nonanone-dependent food leaving’, that assesses food leaving during exposure to the repellent 2-nonanone. Wild type N2 *C. elegans* were examined as the control in comparison to other *Caenorhabditis* species, including, *Caenorhabditis remanei* (C. remanei), *Caenorhabditis nouraguensis* (C. nouraguensis), and *Caenorhabditis portoensis* (C. portoensis).

Wild type N2 *C. elegans* in the lab are able to leave *E. coli* OP50 food lawns efficiently when exposed to undiluted 2-nonanone repellent (Harris et al., 2019, Fig. 1A-B). In each experiment, we compared each nematode species to wild type *C. elegans* Bristol N2 (*Caenorhabditis* Genetics Center, CGC). A *Caenorhabditis* species, *C. nouraguensis* (*Caenorhabditis* sp.17) left an *E. coli* OP50 food lawn during exposure to 2-nonanone similar to wild type N2 *C. elegans*, suggesting *C. nouraguensis* performs 2-nonanone-dependent food leaving with the same food leaving dynamics as wild type *C. elegans* N2 strain (Fig. 1B, Felix MA et al., 2014). Interestingly, we then identified a species that left the food lawn significantly slower than wild type N2 *C. elegans*. The nematode, *C. portoensis* left the *E. coli* OP50 food lawn significantly slower during exposure to 2-nonanone (Fig. 1C). This nematode showed normal movement to the edge of the food lawn furthest away from the 2-nonanone, but remained at the edge of the food lawn for a significantly longer period without leaving the food (Fig. 1C, Felix MA, 2014). In addition to finding nematode species that show delayed food leaving compared to wild type N2 *C. elegans*, we also identified a nematode species that showed enhanced 2-nonanone-dependent food leaving. The nematode species, *C. remanei*, left *E. coli* OP50 food much faster during exposure to 2-nonanone (Fig. 1D, Sudhaus et al., 1974). We also confirmed that for the enhanced food leaving phenotype seen in *C. remanei* was not due to enhancement in sensitivity to 2-nonanone, using the standard 2-nonanone avoidance assay. We found that *C. remanei* showed no enhanced avoidance to 2-nonanone across a time course of 60 minutes (15, 30, 45, 60) (Troemel et al., 1997, Fig. 1H). This suggests that nematodes, such as, *C. remanei* may exhibit differences in food leaving during repellent exposure due to potentially different response to food signals from the food lawn, during the decision to leave food and not due to differences in sensitivity to the 2-nonanone repellent that drives food leaving. It will be interesting to examine whether delayed food leaving observed in *C. portoensis* is related to differences in 2-nonanone sensitivity.

In addition, we have identified that the delayed and enhanced food leaving seen in *C. portoensis* and *C. remanei*, respectively, during exposure to 2-nonanone was also evident when exposing these two nematode species to a second odor repellent, undiluted benzaldehyde, that at high concentrations is also repulsive and drives food leaving in wild type N2 *C. elegans* (Fig. 1E and 1F, Harris et al., 2019, Troemel et al., 1995; Luo et al., 2008). Suggesting, the delayed and enhanced food leaving differences observed are general to multiple repellents. Finally, we examined changing food types during the assay to determine if we observed similar 2-nonanone-dependent food leaving behavior using different food patches. We show that using a *Bacillus megaterium* food patch instead of *E. coli* OP50 patch did not affect the enhanced food leaving in *C. remanei*, suggesting this faster food leaving behavior during exposure to 2-nonanone is general to multiple food types (Fig. 1G). Overall, we show an experiment that establishes variation between nematodes when measuring decision-making to leave a food patch during responses to conflicting cues, such as in this example, food and a repulsive cue, 2-nonanone. This provides an avenue to further examine differences in nematode behavior when exposing the worms to complex sets of attractive and repulsive cues.

**Methods**

Request a detailed protocol

All worms were cultivated under the standard conditions (Brenner et al., 1974). Wild type (N2) hermaphrodites were used in this experiment. Wild type and other *Caenorhabditis* species were prepared in the presence of food (*E. coli* OP50) and staged to use young adult worms for all behavior assays.

**Multi-sensory Behavioral Assay:**

Wild type worms (N2) and all other strains in this study were assayed as previously described (Harris et al., 2019). For all worms, *E.coli* OP50 was used as the food source. For preparation of assay plates, the NGM (Nematode Growth Medium) agar was made and poured into small assay plates (6 cm) and then allowed to cool and solidify (using standard NGM plate preparation protocol). After two days, a liquid suspension of *E. coli* OP50 was prepared; 40 mL of NGM media and *E. coli* OP50 colonies were added to the NGM media, this media was then placed at 26°C overnight. The next morning, the NGM media (*E. coli* OP50 culture) was centrifuged at 3500 rpm for approximately 15 minutes. 35 mL of the NGM was removed and discarded. The *E. coli* OP50 pellet was then re-suspended in 5 mL of NGM through mixing on a table top vortex and then 55 µL of this *E. coli* OP50 culture was then added to the center of an NGM agar assay plate (as seen in Fig. 1A, Schematic of behavioral assay). All worms were grown at 20-23°C on NGM plates that contained a lawn of *E. coli* OP50. For each assay, a population of young adult worms, 25-30 wild type (N2) and different nematodes were placed on separate NGM plates on the bacterial food source (*E. coli*, 55 µL). 2-nonanone (100%, 1µL) was then placed approximately two worm lengths from the food source. All nematodes examined were recorded every 5 minutes to
determine how many worms have left the E. coli OP50 food lawn, until completion at 45 minutes. Food leaving during 2-nonanone exposure was defined as the number of worms that left the food lawn (not present on the food at each 5-minute time point across 45 minutes). For all benzaldehyde experiments, assays were performed in identical conditions to 2-nonanone-dependent food leaving assays. This shows the number of worms that have left the E. coli OP50 food lawn over 45 minutes. Assays with Bacillus megaterium food lawns used identical assay conditions and worm rearing, and wild type N2 C. elegans were used as a control for each condition tested. For data analysis, we analyzed the Mean ± SEM, Student’s t-test, * p ≤ 0.05, **p ≤ 0.01, ***p ≤ 0.001.

2-nonanone avoidance assay

To examine the avoidance of 2-nonanone, chemotaxis assays were performed essentially as previously described (Troemel et al., 1997). Briefly, animals were placed in the center of a square plate that was divided into sectors A, B, C, D, E and F and 2 drops of 1 µl of undiluted 2-nonanone was added to one side and 2 drops of 1 µl ethanol was added to the opposite side of the plate as a control. Approximately 100 worms were used in each assay. 2-nonanone avoidance was analyzed by counting the number of worms in the sectors A-B, C-D, and E-F with E-F being furthest away from the 2-nonanone point sources. The avoidance index was calculated as the number of animals in sectors A and B (at 2-nonanone) minus the number of animals in the sectors E and F (at control) and normalized with the total number of animals in all 6 sectors on plate (Avoidance Index between 0 and -1.0). For all data analysis, a student’s t test was performed when comparing wild type N2 C. elegans hermaphrodite worms to any Caenorhabditis nematode species tested on the same day in parallel conditions. Mean ± SEM, Student’s t-test, * p ≤ 0.05, p ≤ 0.01**, p ≤ 0.001***

Reagents

Chemicals used in this study

All chemicals used in this study, including, 2-nonanone (CasRN: 821-55-6) and benzaldehyde (CasRN: 100-52-7) were purchased from Sigma Aldrich.

List of worm strains and bacterial strains used in these experiments

1) N2 Caenorhabditis elegans Bristol wild type worms were purchased from CGC
2) E.coli OP50 strain purchased from CGC
3) DA1880 Bacillus megaterium purchased from CGC
4) PB219 Caenorhabditis remanei was purchased from CGC
5) EG4788 Caenorhabditis portoensis was purchased from CGC
6) JU1825 Caenorhabditis nouraguensis was purchased from CGC

All strains were provided by the CGC(Caenorhabditis Genetics Center) at the University of Minnesota, which is funded by NIH Office of Research Infrastructure Programs (P40 OD010440).

Acknowledgments: We thank undergraduate lab members of the Harris lab for reading and helpful discussion about the data for relevant experiments. This project was planned and carried out by undergraduate researchers under supervision of the principal investigator of the lab(Dr. Gareth Harris). We thank CGC for strains for this project. We also thank our Biology program Instructional Support Technicians Catherine Hutchinson and Mike Mahoney for their diligent efforts with reagent ordering and helping with aspects of our lab.

References


**Funding:** Laboratory start-up funds were used for this project (PI: Gareth Harris), Biology Program, CSUCI.

**Author Contributions:** Renae Ellis: Formal analysis, Investigation, Methodology, Writing - original draft, Writing - review and editing. Gareth Harris: Formal analysis, Methodology, Writing - review and editing, Data curation, Funding acquisition, Investigation, Supervision, Conceptualization, Resources.

**Reviewed By:** Anonymous

**History:** Received August 3, 2020  Revision received November 13, 2020  Accepted November 17, 2020  Published November 28, 2020

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**Citation:** Ellis, R; Harris, G (2020). Variation between nematodes in a multi-sensory behavioral assay. microPublication Biology. https://doi.org/10.17912/micropub.biology.000330