Analysis of extracellular matrix composition in the visceral muscles of Nidogen mutant larvae in Drosophila

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Figure 1: Distribution of extracellular matrix (ECM) proteins in the visceral midgut muscles of Ndg mutant larvae.

(A) Schematic illustration of ECM core components. Laminin heterotrimers are anchored to cell surface receptors and are organized as a network building ternary nodes. The Laminin A and Laminin Wb heterotrimers are composed of either the α-subunit Laminin A (LanA, red) or the α-subunit Laminin Wb (LanWb, red), the β-subunit Laminin B1 (LanB1, dark red) and the γ-subunit Laminin B2 (LanB2, light red). Collateral linkage between the ECM networks is mediated by Perlecan (Pcan, yellow; encoded by the trol gene) as well as Nidogen (Ndg, blue), that binds to Laminin, Perlecan and Collagen IV. The Collagen IV triple helix is composed of two α1-subunits (Collagen IV α1, Col4α1, dark green) and one α2-subunit (Viking, Vkg, light green). Secreted protein, acidic, cysteine-rich (SPARC, purple) binds to the Collagen IV.
network. (B) Schematic illustration of the larval midgut visceral muscle morphology. Circular muscles highlighted in light orange and overlaying longitudinal muscles in dark orange. (C, D) Nidogen antibody staining in control (white, C) and NdgΔ1.4 mutant larvae (D). (E, F) Laminin A and (G, H) Laminin Wing blister antibody staining in control (white, E and G) and NdgΔ1.4 mutant larvae (F and H). (I, J) GFP antibody staining in control (trol::GFP, I) and Ndg mutant (trol::GFP; NdgΔ1.4, J), (K, L) as well as in control (vkg::GFP, K) and Ndg mutant larvae (vkg::GFP, NdgΔ1.4, L). (M, N) SPARC antibody staining in control (white, M) and NdgΔ1.4 mutant larvae (N). Scale bars = 50 μm.

Description
A group of highly conserved extracellular matrix (ECM) proteins, which are enriched in basement membranes across all metazoa, form the so-called ‘basement membrane tool kit’ (Hynes, 2012; Hynes and Zhao, 2000). These important components are organized in two layers connected to each other via the linker proteins Perlecan and Nidogen (Fig. 1A). The inner layer is formed by a self-assembling network of Laminin heterotrimers, consisting of an α-, β- and γ-subunit each. The outer network is composed of Collagen IV, which builds a triple helix with two α1-subunits and one α2-subunit. In Drosophila melanogaster a small set of genes encode for these components. Four genes code for Laminin subunits: two genes for α-subunits, (Laminin A, LanA, and wing blister, wb, referred to herein as ‘LanWb’), and only one gene encodes for a β- (Laminin B1, LanB1) and one gene encodes for an γ-subunit (Laminin B2, LanB2). Furthermore, one gene encodes for the Collagen IV subunit α1 (Collagen IV α1, Col4α1) and α2 (viking, vkg) as well as for the linker proteins Perlecan (terribly reduced optic lobes, trol) and Nidogen (Ndg). Ndg connects Laminin to the Collagen IV network via binding of Perlecan and Collagen IV to the Laminin γ-subunit, and has been suggested to play an essential role in ECM assembly (Fig. 1A; Fox et al., 1991; Hopf et al., 2001a; Mann et al., 1989; Reinhardt et al., 1993). However, in contrast to mutants of other basement membrane components in Drosophila, Ndg mutants are viable and seem not to play a role in general ECM assembly (Dai et al., 2018; Wolfstetter et al., 2019), whereas further ultrastructural analyses reveal a disrupted ECM of larval visceral muscles (Wolfstetter et al., 2019). Whether this ultrastructural alterations in the larval visceral muscles of Ndg mutants correspond to a change in distribution or assembly of other basement membrane components is not known, so we examine the protein distribution of the main ECM components in Ndg mutants in this study.

Here, we analysed the relevance of Ndg for ECM assembly in the visceral muscles of third instar larvae and compared basement membrane protein localization in visceral muscles of amorphic Ndg mutants to controls through labelling with specific antibodies and GFP exon trap lines. As expected, Ndg is evenly distributed across all visceral muscles in controls (Fig. 1C) whereas Ndg mutants do not show this signal (Fig. 1D), although the visceral muscles are properly formed as detected by control staining of F-actin (data not shown). The assembly of Laminin heterotrimers is essential for proper incorporation of other ECM components like Ndg (Wolfstetter and Holz, 2012; Wolfstetter et al., 2019). Since visceral muscles express both possible Laminin heterotrimer and there could be some functional redundancy between them (Martin et al., 1999; Wolfstetter and Holz, 2012; Yarnitzky and Volk, 1995), we studied the distribution of both Laminin α-subunits (LanA in Fig. 1E and F and LanWb in Fig. 1G and H). Congruently with the role of the Laminin network for basement membrane initiation (Hohenester and Yurchenco, 2013), loss of Ndg neither affect Laminin A nor the Wing blister heterotrimer deposition in the larval visceral muscles in general (Fig. 1E-H). The linker protein Pcan is supposed to act as collateral linker between the Laminin and the Collagen networks, which could be explained by a redundant function of Pcan and Ndg (Fig. 1A). This model is comparable to the role of the Laminin network for basement membrane initiation (Hohenester and Yurchenco, 2013), loss of Ndg neither affect Laminin A nor the Wing blister heterotrimer deposition in the larval visceral muscles in general (Fig. 1E-H). The linker protein Pcan is supposed to act as collateral linker between the Laminin and the Collagen networks, which could be explained by a redundant function of Pcan and Ndg (Fig. 1A).

Our data confirm that the basic composition of the examined ECM proteins is not affected by loss of Ndg and that Ndg therefore does not appear to be the essential single player for the connection of the different basement membrane layers.

Methods
Immunofluorescence staining
Antibody staining of Drosophila wandering third instar larvae was performed as described by Müller, (2008) and modified as described in Wolfstetter et al. (2019). The following primary antibodies were used: rabbit anti-green fluorescent protein (GFP, 1:500, Abcam, ab290), guinea pig anti-Laminin A (LanA, 1:500; Harpaz and Volk, 2012), rabbit anti-Wing blister (LanWb, 1:100; Martin et al., 1999), rabbit anti-Nidogen (Ndg, 1:1.000; Wolfstetter et al., 2009), rabbit anti-SPARC (SPARC, 1:500; Martinek et al., 2002), Cy3 conjugated goat IgG anti-guinea pig (1:200, Dianova) and DyLight 488.
conjugated goat anti-rabbit secondary antibody (1:1000; Vector Laboratories). Nonspecific binding sites were blocked with 5% goat serum. Tissues were embedded in Fluoromount G (Southern Biotech) and imaged by confocal microscopy (Leica TCS SP2) with same laser intensity for each experiment.

Fly stocks and genetics

Flies were grown under standard conditions (Ashburner, 1989) and crosses were performed at 25 °C. As control stocks we used white1118 (w118, FBal0018186), trol::GFP (FBal0243609) and vkg::GFP (FBal0191275) as exon trap lines (Morin et al., 2001). For analyses in Ndg mutant background, NdgΔ1.4 (Wolfstetter et al., 2019, FBal0346270), trol::GFP; NdgΔ1.4 and vkg::GFP, NdgΔ1.4 were used.

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References


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