

Two Deletion Alleles in the *C. elegans mir-49* gene.

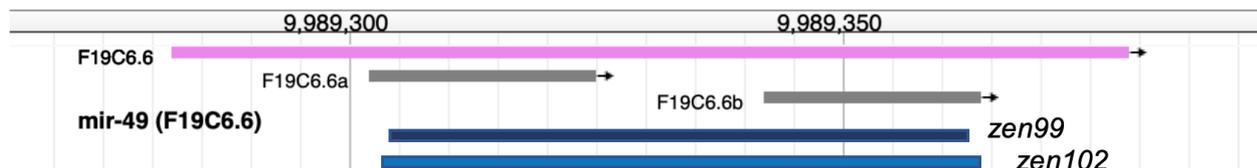
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*These authors contributed equally.

A



B

wt
zen99
F19C6.6
mir-49-5p
mir-49-3p

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CAGATGCTAGTGTACTTTTGAARAAAGACCACCGTCCGCGAGTTTGTGTGATGTGCTCCAGCARTCATGAGTCTGARGCACCACGAGAGCTGCAGATGGAGGTTCTGATTTTCGARCAGGTTTCAGACARAGTCG
CAGATGCTAGTGTACTTTTGAARAAAGACCACCGTCCGCGAG:::ATGGAGGTTCTGATTTTCGARCAGGTTTCAGACARAGTCG
AGTGTACTTTTGAARAAAGACCACCGTCCGCGAGTTTGTGTGATGTGCTCCAGCARTCATGAGTCTGARGCACCACGAGAGCTGCAGATGGAGGTTCTGATTTTCGARCAGGTTTCAGAC
CGCAGTTTGTGTGATGTGCTCC
AAGCACCACGAGAGCTGCAG
  
```

C

wt
zen102
F19C6.6
mir-49-5p
mir-49-3p

```

TCAGATGCTAGTGTACTTTTGAARAAAGACCACCGTCCGCGAGTTTGTGTGATGTGCTCCAGCARTCATGAGTCTGARGCACCACGAGAGCTGCAGATGGAGGTTCTGATTTTCGARCAGGTTTCAGACARAGTCG
TCAGATGCTAGTGTACTTTTGAARAAAGACCACCGTCCGCGA:::TGGAGGTTCTGATTTTCGARCAGGTTTCAGACARAGTCG
AGTGTACTTTTGAARAAAGACCACCGTCCGCGAGTTTGTGTGATGTGCTCCAGCARTCATGAGTCTGARGCACCACGAGAGCTGCAGATGGAGGTTCTGATTTTCGARCAGGTTTCAGAC
CGCAGTTTGTGTGATGTGCTCC
AAGCACCACGAGAGCTGCAG
  
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Figure 1: (A) A schematic of the *mir-49* locus and the location of the newly generated *mir-49(zen99)* and *mir-49(zen102)* deletion alleles. (B) *zen99* removes 56 base pairs from the *mir-49* precursor. (C) *zen102* removes 58 base pairs from the *mir-49* precursor.

Description

MicroRNAs (miRNAs) are small, non-coding RNAs that post-transcriptionally repress gene expression (Gebert and MacRae, 2018). While many miRNA genes and their families have been analyzed for function (Miska *et al.* 2007, Alvarez-Saavedra and Horvitz 2010), there are microRNA genes for which loss of function alleles have not yet been generated. There are no available alleles for the *C. elegans mir-49* gene.

Using CRISPR-Cas9 genome editing, we generated two deletion alleles, *zen99* and *zen102*, that disrupt the *C. elegans mir-49* gene (Fig 1A). *mir-49(zen99)* and *mir-49(zen102)* delete 56 base pairs and 58 base pairs from the *mir-49* locus, respectively (Fig 1B and Fig 1C). Each deletion nearly completely removes both strands generated by the *mir-49* locus, *mir-49-3p* and *mir-49-5p*. Both *mir-49* alleles are homozygous viable and appear to be superficially wild type. Careful phenotypic analysis will be important to characterize the effects of the two *mir-49* deletions.

Methods

To generate the *mir-49* deletion alleles, N2 animals were injected with the CRISPR-Cas9 components as an RNA-protein complex (Paix *et al.* 2015). The following components were used: Alt-R Cas9 (IDT, cat# 1081058) loaded with *mir-49* crRNAs (IDT, custom) (*mir-49* crRNA1 sequence: 5'-GAGCACATCACAACTG-3', *mir-49* crRNA2 sequence: 5'-GCACCACGAGAAGCTGCAGA-3'), *dpy-10* targeting guide RNA (IDT, custom) (5'-GCUACCAUAGGCACCACGAG-3', Arribere *et al.* 2014) and tracer RNA (IDT, cat# 1072532) (AGCAUAGCAAGUUAUUAAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGCUUU).

Briefly, to load the Alt-R Cas9, the following mixture was incubated at 37°C for 15 minutes: 0.5µL of Alt-R Cas9, 2.4µL of tracrRNA (0.4µg/µL), 0.8µL of *mir-49* crRNA1 (0.4µg/µL), 0.8µL of *mir-49* crRNA2 (0.4µg/µL), 1.3 µL of *dpy-10* crRNA (0.1µg/µL), 1µL IDT annealing buffer (provided with Alt-R Cas9), and 3.2µL of water. Following the incubation, the mixture was spun for 2 minutes at top speed (~10,000rpm). The progeny of the injected animals was first screened for the presence of dumpy worms to identify parents positive for Cas9 activity (Arribere *et al.* 2014). F1 offspring of the Cas9-positive parents were then genotyped for the presence of potential *mir-49* deletions using the following primers: mir-

49.for1 (5'-AGGCACCACCACTTACCATTTCAT-3') and mir-49.rev1 (5'-GATGACTTACAGTCGCGTCTT-3'), which generate a wild type product of ~430 bps. Independent *mir-49* deletions were identified, homozygosed, and sequenced. The resultant strains, UY264 (*mir-49(zen99)*) and UY267 (*mir-49(zen102)*) were not outcrossed, but appear to be free of background *dpy-10* mutations. Sequencing was repeated in the next generation to ensure the stability of the generated alleles.

Reagents

UY264 *mir-49(zen99)* and UY267 *mir-49(zen102)* are available upon request.

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References

- Gebert LFR and Macrae IJ (2019). Regulation of microRNA function in animals. *NatureReviews Molecular Cell Biology*, 20(1), 21–37. DOI: 10.1038/s41580-018-0045-7 | PMID: 30108335.
- Miska EA, Alvarez-Saavedra E, Abbott AL, Lau NC, Hellman AB, McGonagle SM, et al. (2007). “Most *Caenorhabditis elegans* microRNAs Are Individually Not Essential for Development or Viability.” *PLoS Genet* 3(12): e215. DOI: 10.1371/journal.pgen.0030215 | PMID: 18085825.
- Alvarez-Saavedra E, Horvitz HR (2010). “Many families of *C. elegans* microRNAs are not essential for development or viability.” *Curr Biol* 20(4):367-73. DOI: 10.1016/j.cub.2009.12.051 | PMID: 20096582.
- Paix A, Folkmann A, Rasoloson D, Seydoux G (2015). “High Efficiency, Homology-Directed Genome Editing in *Caenorhabditis elegans* Using CRISPR-Cas9 Ribonucleoprotein Complexes.” *GENETICS* 201(1): 47-54. DOI: 10.1534/genetics.115.179382 | PMID: 26187122.
- Arribere JA, Bell RT, Fu BX, Artiles KL, Hartman PS, Fire AZ (2014). “Efficient Marker-Free Recovery of Custom Genetic Modifications with CRISPR/Cas9 in *Caenorhabditis elegans*.” *GENETICS* 198(3): 837-846. DOI: 10.1534/genetics.114.169730 | PMID: 25161212.

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