Calx, a sodium/calcium exchanger, may affect lifespan in *Drosophila melanogaster*

Jung-Wan Mok, Hyunglok Chung and Kwang-Wook Choi

1Department of Biological Sciences, Korea Advanced Institute of Science and Technology, Daejeon 34141, Korea
2Current address: Department of Molecular and Human Genetics, Baylor College of Medicine, Houston, TX 77030, USA
§To whom correspondence should be addressed: kchoi100@kaist.ac.kr

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**Figure 1:** A. Calx protein expression profile in *Calx* homozygous mutants. B. Lifespan assay at 25°C. C. Lifespan assay at 29°C. D. Statistics for (B) and (C).

**Description**

Calcium homeostasis is essential for normal body function. Calx is a *Drosophila* homolog of the mammalian sodium/calcium exchanger (NCX) involved in the regulation of intracellular calcium (*Ca^{2+}*). A major function of Calx is to export excess *Ca^{2+}* to the outside of the cell when intracellular *Ca^{2+}* level is elevated (Hryshko et al. 1996). Genetic studies in *Drosophila* have shown that *Calx* mutants can develop to adult flies, even though *Calx* is expressed throughout the developmental stages (Wang et al. 2005). However, *Ca^{2+}* efflux mediated by Calx is crucial for maintaining the cellular *Ca^{2+}* homeostasis in light-activated sensory neurons (Wu et al. 2011). In addition to its role for phototransduction, Calx is also required for preventing light-induced retinal degeneration caused by *Ca^{2+}* overload (Wang et al. 2005).

Thus far, Calx function has been extensively studied in the retinal cells. However, expression of *Calx* gene is not restricted to the eye, and potential functions of Calx in other tissues are largely unknown. Interestingly, disrupted *Ca^{2+}* homeostasis has been implicated in aging and is regarded as one of the biomarkers of aging (Foster and Kumar 2002). Although mammalian NCXs have been implicated in aging of tissues and organs (Gomez-Villafuertes et al. 2007; Zhang et al.)
To determine whether Calx is required for normal lifespan, we carried out genetic analysis using two Calx mutants, CalxA and CalxB. It has been shown that homozygous CalxA and CalxB mutant flies are viable (Wang et al. 2005). CalxA mutants have a point mutation (T822I) but show relatively normal level of protein expression in adult head. In contrast, CalxB is a regulatory mutation that causes reduced protein expression (Wang et al. 2005). To check whether such mutant effects can also be seen in non-head tissues, we examined Calx protein expression in whole adult fly body. It has been reported that Calx is expressed as a single 110 kD protein in adult head and body (Wang et al., 2005). w1118 flies used as wild-type control showed a major band at 110 kD and a weaker band at 120 kD (Fig. 1A). Consistent with earlier studies (Wang et al. 2005), we found a strong reduction in the 110 kD band in CalxB mutant, while CalxA mutant showed a nearly indistinguishable pattern from wild-type. CalxB mutant showed severe reduction in both 110 and 120 kD bands, suggesting that Calx exists at least in two different forms in adult fly.

Next, we measured lifespan of wild-type and Calx males. We used isogenic w1118 as control since it was previously used as control for analyzing Calx mutant phenotypes in phototransduction (Wang et al. 2005). In our lifespan assay at 25 °C, both CalxA and CalxB mutants showed considerable reduction in the survival rate compared with the wild-type control (Fig. 1B). Median lifespan of CalxA and CalxB mutants was 24 and 21.5 days, respectively, whereas the median for wild-type was about 36 days (Fig. 1D). We also examined survival rates of Calx mutants at 29 °C (Fig. 1C). At this high temperature, wild-type flies showed a lower survival rate with median lifespan of 21.5 days. Under this condition, CalxA mutant flies had a significantly shorter median lifespan than CalxB mutant (12.5 days for CalxA; 17.7 days for CalxB) (Fig. 1D). Since CalxA encodes a mutated protein, it is possible that the mutant Calx protein might be more unstable at 29 °C, causing more severe phenotype in longevity. Alternatively, CalxA mutant cells may require a higher rate of calcium extrusion at the upper extreme of their viable temperature range than their more normal growth temperature (25 °C). We also checked the lifespan of transheterozygotes for CalxA and CalxB mutations (CalxA/CalxB). Trans-heterozygous flies also showed significantly reduced lifespan compared with wild-type, indicating that CalxA and CalxB fail to complement the longevity phenotype. Consistent with the hypomorphic nature of the CalxB allele, trans-heterozygous flies showed an intermediate lifespan between CalxA and CalxB mutants (Fig. 1C, D). Taken together, our data suggest that the Calx mutations are responsible for the observed phenotype of shorter longevity. These results are preliminary, and to rule out potential genetic background effects, additional controls should be included in the future.

This study raises a possibility that Calx might be required for normal lifespan. The Ca2+ influx channel TRP, which has an opposite function to Calx, is crucial for phototransduction (Montell 2005; Wang et al. 2005). The trp gene has a paralog, trp-like (trpl) (Phillips et al. 1992). Although there is no strict paralog of Calx in Drosophila, Nckx30C is a distant relative of Calx (Haug-Collet et al. 1999). It remains to be tested whether these related genes are also required for normal lifespan. Importantly, retinal degeneration by loss of TRP or constitutive TRP function can be suppressed by defects or overexpression of Calx, respectively. Further studies are necessary to see whether lifespan is dependent on the functional relationship between Calx and TRP in the intracellular Ca2+ regulation, although such genetic analysis may be complicated by the existence of related genes like trpl and Nckx30C. It is also an intriguing question whether NCX homologs of Calx play a role in lifespan of mammals.

**Methods**

**Western Blot** – For western blot experiment, 10 adult male flies for each genotype (3 days old virgin) were used for protein extraction in 200μl of SDS sample buffer. After boiling for 5 minutes at 94 °C, samples were centrifuged at 12000g for 10 minutes. Supernatants were loaded on 10% polyacrylamide gel. Fractionated proteins were transferred by western blotting for immunostaining (Calx antibody 1:1000, kind gift from Dr. Craig Montell).

**Lifespan assay** – For lifespan assay, one hundred adult male flies of each genotype (1 day old, virgin) were collected by minimal exposure to carbon dioxide gas. Flies were raised in food vials (10 males/vial) and transferred to new vials every 2-3 days. Number of dead flies was recorded every day. Statistical significance of the data from lifespan assay was determined by P-value evaluation from two-tailed T test.

**Reagents**

Fly strains – Isogenized w1118 (#5905), CalxA (#24496) and CalxB (#24497) mutants were kindly provided by the Bloomington Drosophila Stock Center.
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References


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